#### PART 1.

CONCORDANCE BETWEEN CANCER IN HUMANS AND IN EXPERIMENTAL ANIMALS

#### CHAPTER 6.

# Anticancer agents: qualitative and quantitative aspects

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#### Introduction

Typically, regulation of human exposure to carcinogenic compounds is based mainly on qualitative considerations for compounds that cause cancer in experimental animals. This approach is based on the old paradigm of using animal models to understand human physiology and pathology; in the regulatory setting, no alternatives to this paradigm have been available specifically in relation to cancer (Maronpot et al., 2004).

Quantitative aspects of carcinogenesis, including estimates of carcinogenic potency in animals and in humans, would have both regulatory and scientific implications. For animals, a systematic assessment

of potency based on bioassay data has been generated from values of TD<sub>50</sub>, i.e. the chronic dose rate (in mg/kg body weight [bw]/day) that is estimated to reduce the proportion of tumour-free animals by 50% (Gold et al., 2005). For humans, exposures to ionizing radiation, occupational carcinogens, and tobacco smoke have been the primary sources of quantitative data on cancer risks, including considerations of dose, duration of exposure, and latency period (i.e. time from exposure to occurrence of cancer) (Breslow and Day, 1987; Moolgavkar et al., 1999; Brenner et al., 2003; Pierce and Vaeth, 2003; Preston et al., 2003, 2004; IARC, 2004). However, because of the complexities of these

various human exposures, comparisons of potency assessments with data from animals have been hampered.

An additional source of quantitative data on cancer risks is the group of patients who are survivors of first primary cancers after treatment with anticancer therapy and who are monitored for treatment-related risks of second primary cancers (Travis, 2006; Travis et al., 2006). Active research on second cancers has been carried out since the 1980s, but because of the increasing numbers of patients enrolled and the extended periods of follow-up, the more recent studies provide the most comprehensive evidence on the magnitude of the effects

**Table 6.1.** Tumour sites and histological types of cancer induced in humans and in rodents after exposure to anticancer agents

Agent	Tumour sites and histological types				
	Humans	Rats	Mice		
Cyclophosphamide	AML	Lymphoma	Lymphoma		
	Bladder cancer	Leukaemia	Acute lymphocytic leukaemia		
		Mammary adenoma	Mammary carcinoma		
		Transitional cell carcinoma of the bladder	Lung cancer		
		Neurogenic sarcoma	Liver cancer		
Chlorambucil	AML	Lymphoma	Lymphoma		
		Myeloid leukaemia	Myeloid leukaemia		
		Mammary carcinoma			
Melphalan	AML	Retroperitoneal sarcoma	Lymphoma		
Thiotepa	Leukaemia	Lymphocytic leukaemia	Lymphoma		
		Uterine sarcoma	Lymphocytic leukaemia		
		Squamous cell cancers of the skin and the ear			
		canal			
AML, acute myeloid le Source: Compiled from					

(Hijiya et al., 2007; Hodgson et al., 2007; Maule et al., 2007; Hemminki et al., 2008; Swerdlow et al., 2011). Although these studies present valuable data on many exposure-related aspects, relevant treatments are seldom based on single agents or single modalities, and individual carcinogens can rarely be singled out. Nevertheless, striking new data from these studies show increased risks of almost all site-specific cancers that emerge during the follow-up period. Such data challenge the "canonical" site-specificity of carcinogenesis. In their review of human carcinogens, Cogliano et al. indicate that no agents classified as carcinogenic to humans (Group 1) are identified as causing prostate cancer (Cogliano et al., 2011). However, some evidence is available. The risk of prostate cancer is significantly increased in survivors of non-Hodgkin lymphoma after chemotherapy, for those diagnosed at age 40-49 years

(Hemminki et al., 2008). Anticancer agents are also used in some other cases, such as for certain autoimmune diseases, but even if a single anticancer agent is given, other medication and the inherent cancer risk of some autoimmune conditions may limit the applicability of the results.

In this chapter, data on anticancer agents from Volume 100A of the IARC Monographs are used to make qualitative comparisons between cancers induced in humans and in experimental animals, with reference to the possible underlying mechanisms. Furthermore, quantitative comparisons of carcinogens with respect to potency in humans and in experimental animals are discussed. This review is limited to anticancer agents for which the evidence of carcinogenicity was considered to be sufficient both in humans and in experimental animals: cyclophosphamide, chlorambucil, melphalan, and thiotepa (IARC, 2012).

Of the anticancer agents included in Volume 100A, the current selection does not include busulfan, 1-(2-chloroethyl)-3-(4-methylcyclohexyl)-1-nitrosourea (methyl-CCNU), treosulfan, and some mixtures of anticancer agents for which evidence is *sufficient* in humans but evidence is *limited* or lacking in animals.

### Therapeutic applications and trends in use

The four anticancer agents discussed here were first used in clinical practice in the 1960s, but since then the clinical indications have been narrowed and their therapeutic use has declined, with the possible exception of cyclophosphamide.

Cyclophosphamide may be used alone for the treatment of several types of cancer, but most often it is administered in combination with other drugs. Diseases for which cyclophosphamide is the recognized treatment include breast cancer, **Table 6.2.** Values of  $TD_{50}$  (the chronic dose rate estimated to reduce the proportion of tumour-free animals by 50%) for anticancer agents in rodents

Agent	TD <sub>50</sub> (mg/kg bw/day)				
	All tumours combined		Haematopoietic malignancies		
	Rats	Mice	Rats	Mice	
Cyclophosphamide	2.2ª	2.8	3.4 <sup>b</sup>	7.9	
Chlorambucil	0.7	0.1	1.6	0.6	
Melphalan	0.6	0.1	0.9	0.5	
Thiotepa	0.04	0.07	0.2	0.2	

bw, body weight.

<sup>a</sup> Bladder tumours in rats: TD<sub>50</sub> = 21 mg/kg bw/day (Gold et al., 1987).

<sup>b</sup> From Gold et al. (1987).

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lymphoma, leukaemia, sarcoma, and ovarian cancer. Cyclophosphamide is also used for treatment of diseases other than cancer, such as nephrotic syndrome and many autoimmune diseases, including Wegener granulomatosis, rheumatoid arthritis, lupus erythematosus, mycosis fungoides, and several forms of vasculitis.

The current clinical use for chlorambucil mainly involves treatment of chronic lymphocytic leukaemia. Chlorambucil may also be used for treatment of non-Hodgkin lymphoma, Waldenström macroglobulinaemia, polycythaemia vera, trophoblastic neoplasms, and ovarian cancer. Chlorambucil has also been applied as an immunosuppressive drug for various autoimmune and inflammatory conditions.

The use of melphalan has declined for treatment of most cancers, but since about 2000 it has been given in high doses to patients with myeloma in combination with autologous stem cell transplantation.

Thiotepa has previously been used in the palliation of a wide variety of neoplastic diseases. It may still be prescribed in intravesical chemotherapy for bladder cancer.

#### **Tumour sites**

Tumour sites and histological types of cancer induced in humans and in rodents by the four anticancer drugs are listed in Table 6.1. In humans, cyclophosphamide causes acute myeloid leukaemia and bladder cancer of undefined histology. Lymphoma, leukaemia, and mammary carcinoma have been detected in rats and mice after administration of cyclophosphamide. In rats, transitional cell carcinoma of the bladder and neurogenic sarcoma have been reported. In mice, cancers of the lung and liver have been detected. Chlorambucil causes acute myeloid leukaemia in humans. Lymphoma and leukaemia have been detected in rats and mice, as well as mammary carcinoma in rats. Melphalan causes acute myeloid leukaemia in humans, retroperitoneal sarcoma in rats, and lymphoma in mice. Thiotepa causes leukaemia in humans and lymphocytic leukaemia in rats and mice. It has been reported to induce uterine sarcoma and squamous cell cancers of the skin and the ear canal in rats and lymphoma in mice.

#### **Carcinogenic potency**

Gold et al. have systematically analysed the carcinogenic potency of compounds tested in animal experiments in the context of the United States National Toxicology Program (Gold et al., 1987, 2005). These data were used by Kaldor et al. to quantify the carcinogenic potency of anticancer agents (Kaldor et al., 1988). With respect to potency, it should be noted that low daily doses producing cancer (i.e. low TD<sub>50</sub> values) indicate high carcinogenic potency.

Data on rats and mice from Kaldor et al. (1988) are collected in Table 6.2. For ease of analysis, results for male and female rodents were averaged. When information was lacking in the paper by Kaldor et al., data were taken from other sources, as indicated. The TD<sub>50</sub> values for all tumours combined are lower than those for haematopoietic malignancies. According to these data, thiotepa is the most potent carcinogen, with TD<sub>50</sub> values of 0.04 mg/kg bw/day in rats and 0.07 mg/kg bw/ day in mice for all tumours combined. Chlorambucil and melphalan **Table 6.3.** Estimated carcinogenic potency (10-year cumulative incidence [%] divided by total dose in grams) in humans of anticancer agents

Agent	Le	Bladder cancer		
	From Kaldor et al. (1988)	<b>Calculated</b> <sup>a</sup>		_
	_	Low dose	High dose	_
Cyclophosphamide	0.28	_	0.04	0.02 <sup>b</sup> , 0.1 <sup>c</sup>
Chlorambucil	4.2, 1.8	16.5	1.4	_
Melphalan	18.7, 3.3	14.1	11.5	-
Thiotepa	_	55.3	3.2	_

for primary ovarian cancer (Kaldor

° Calculated from Travis et al. (1995).

are equally potent as carcinogens, whereas cyclophosphamide is weaker by approximately an order of magnitude. After treatment of rats with cyclophosphamide, bladder cancer was detected with a  $TD_{50}$  of 21 mg/kg bw/day, i.e. an order of magnitude lower than the value for all tumours combined (Gold et al., 1987).

The measure of potency used by Kaldor et al. was the 10-year cumulative incidence of leukaemia (a percentage) divided by the total administered dose in grams; thus, a large number indicates high potency (Kaldor et al., 1988). These data are shown in Table 6.3. Information was lacking for thiotepa, but of the remaining compounds melphalan was the most potent, with values of 18.7 and 3.3 from two separate studies. Chlorambucil showed an intermediate potency, which was an order of magnitude higher than that of cyclophosphamide.

More recent data were added to Table 6.3 from studies in which the anticancer agent was the principal drug used and no radiotherapy was applied. Kaldor et al. published a multinational study of secondary leukaemias in women after treatment

et al., 1990). The potency according to Kaldor et al. was calculated from the cumulative baseline incidence of leukaemia in Sweden of 0.2 per 10 years, multiplied by the relative risk given in the relevant paper; the product was then divided by the median doses cited for the low dose and the high dose (Kaldor et al., 1988). The low and high doses differed widely, and the calculated potency values were clearly higher for the low dose than for the high dose. Thiotepa and melphalan were the most potent drugs, followed by chlorambucil and the much weaker cyclophosphamide. The potency of cyclophosphamide to induce bladder cancer was also calculated, according to the data from two studies; in the first study, bladder cancer was diagnosed in women after treatment for ovarian cancer (Kaldor et al., 1995), and in the second study, bladder cancer was diagnosed in survivors of non-Hodgkin lymphoma (Travis et al., 1995). The use of a cumulative baseline incidence of bladder cancer in women of 0.4 per 10 years for the data of the first study and a sex-adjusted incidence of 0.9 per 10 years for the second study resulted in potency values of 0.02 and 0.1, respectively (Table 6.3). Thus, the potency of cyclophosphamide determined in relation to bladder cancer was lower than its potency in the haematopoietic system. A similar outcome was evident in the rodent studies (Table 6.2).

#### **Mechanisms of action**

Cyclophosphamide activated is through cytochrome P450а mediated reaction to yield phosphoramide mustard and acrolein, both of which can bind to DNA. Phosphoramide mustard undergoes rapid dephosphoramidation, which in neutral in vitro conditions proceeds with a half-life of 8 minutes, resulting in the formation of nornitrogen mustard (Hemminki, 1985). Because most of the metabolic activation of cyclophosphamide takes place in the liver, it seems likely that a considerable proportion of DNA binding in peripheral tissues is in fact mediated by nornitrogen mustard (Hemminki, 1985). As summarized in Volume 100A of the IARC Monographs, cyclophosphamide has several endpoints indicative of genotoxic effects in humans, including DNA damage as measured by the comet assay, mutations at the *HPRT* locus, and sister chromatid exchange. Historically, cyclophosphamide has been included in several genetic structure–activity studies (Vogel et al., 1996, 1998).

Chlorambucil and melphalan are direct-acting derivatives of nitrogen mustard, and thiotepa is a direct-acting trifunctional derivative of aziridine. These compounds bind to DNA and give a positive response in a wide spectrum of assays for genomic injury, including tests for cytogenetic damage, specifically as indicated by chromosomal aberrations and sister chromatid exchange in patients. These drugs have also been included in several genetic structure– activity studies (Vogel et al., 1996, 1998).

#### Conclusions

For the anticancer drugs cyclophosphamide, chlorambucil, melphalan, and thiotepa, the data summarized in this chapter show that the target sites for which there is *sufficient evidence* of carcinogenicity are generally similar in rodents and humans, particularly for bladder cancer induced by cyclophosphamide.

Anticancer agents allow unique comparisons of carcinogenic potency among species, because the doses administered to humans and animals are known. Cancer treatment has become increasingly multimodal and involves the use of multiple drugs; this makes it difficult to single out individual agents. Also, survival rates have risen and the probability of detecting second tumours has increased. It is unclear why there is not more research activity to follow up other patient groups who receive anticancer agents, such as patients with autoimmune diseases. The current potency data for four anticancer drugs suggest that the TD<sub>50</sub> values for rats and mice are reasonably homogeneous and consistent. As a carcinogen, cyclophosphamide was the least potent and thiotepa was the most potent agent in any of the rodent models analysed. In humans, cyclophosphamide was the least potent and thiotepa and melphalan were the most potent compounds to induce secondary cancers.

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## References

Brenner DJ, Doll R, Goodhead DT, Hall EJ, Land CE, Little JB, et al. (2003). Cancer risks attributable to low doses of ionizing radiation: assessing what we really know. Proc Natl Acad Sci U S A. 100(24):13761–6. http://dx.doi.org/10.1073/pnas.2235592100 PMID:14610281

Breslow NE, Day NE (1987). Statistical methods in cancer research. Volume II – The design and analysis of cohort studies. Lyon, France International Agency for Research on Cancer (IARC Scientific Publications, No. 82).

Cogliano VJ, Baan R, Straif K, Grosse Y, Lauby-Secretan B, El Ghissassi F, et al. (2011). Preventable exposures associated with human cancers. J Natl Cancer Inst. 103(24):1827–39. <u>http://dx.doi.org/10.1093/</u> inci/djr483 PMID:22158127

Gold LS, Manley NB, Slone TH, Rohrbach L, Garfinkel GB (2005). Supplement to the Carcinogenic Potency Database (CPDB): results of animal bioassays published in the general literature through 1997 and by the National Toxicology Program in 1997–1998. Toxicol Sci. 85(2):747–808. <u>http://dx.doi.org/10.1093/toxsci/kfi161 PMID:15800034</u>

Gold LS, Slone TH, Backman GM, Magaw R, Da Costa M, Lopipero P, et al. (1987). Second chronological supplement to the Carcinogenic Potency Database: standardized results of animal bioassays published through December 1984 and by the National Toxicology Program through May 1986. Environ Health Perspect. 74:237–329. <u>http://dx.doi.org/10.1289/ehp.8774237</u> PMID:3691431

Hemminki K (1985). Binding of metabolites of cyclophosphamide to DNA in a rat liver microsomal system and in vivo in mice. Cancer Res. 45(9):4237–43. <u>PMID:4028012</u>

Hemminki K, Lenner P, Sundquist J, Bermejo JL (2008). Risk of subsequent solid tumors after non-Hodgkin's lymphoma: effect of diagnostic age and time since diagnosis. J Clin Oncol. 26(11):1850–7. http://dx.doi.org/10.1200/JCO.2007.14.6068 PMID:18347006

Hijiya N, Hudson MM, Lensing S, Zacher M, Onciu M, Behm FG, et al. (2007). Cumulative incidence of secondary neoplasms as a first event after childhood acute lymphoblastic leukemia. JAMA. 297(11):1207–15. <u>http://</u> <u>dx.doi.org/10.1001/jama.297.11.1207</u> <u>PMID:17374815</u>

Hodgson DC, Gilbert ES, Dores GM, Schonfeld SJ, Lynch CF, Storm H, et al. (2007). Longterm solid cancer risk among 5-year survivors of Hodgkin's lymphoma. J Clin Oncol. 25(12):1489–97. <u>http://dx.doi.org/10.1200/</u> JCO.2006.09.0936 PMID:17372278 IARC (2004). Tobacco smoke and involuntary smoking. IARC Monogr Eval Carcinog Risks Hum. 83:1–1438. <u>PMID:15285078</u>. Available from: <u>http://publications.iarc.fr/101</u>.

IARC (2012). Pharmaceuticals. IARC Monogr Eval Carcinog Risks Hum. 100A:1–437. <u>PMID:23189749</u>. Available from: <u>http://</u> publications.iarc.fr/118.

Kaldor JM, Day NE, Hemminki K (1988). Quantifying the carcinogenicity of antineoplastic drugs. Eur J Cancer Clin Oncol. 24(4):703–11. <u>http://dx.doi.org/10.1016/0277-5379(88)90302-1 PMID:3383970</u>

Kaldor JM, Day NE, Kittelmann B, Pettersson F, Langmark F, Pedersen D, et al. (1995). Bladder tumours following chemotherapy and radiotherapy for ovarian cancer: a case-control study. Int J Cancer. 63(1):1–6. http://dx.doi.org/10.1002/ijc.2910630102 PMID:7558434

Kaldor JM, Day NE, Pettersson F, Clarke EA, Pedersen D, Mehnert W, et al. (1990). Leukemia following chemotherapy for ovarian cancer. N Engl J Med. 322(1):1–6. <u>http://</u><u>dx.doi.org/10.1056/NEJM199001043220101</u> PMID:2104664

Maronpot RR, Flake G, Huff J (2004). Relevance of animal carcinogenesis findings to human cancer predictions and prevention. Toxicol Pathol. 32(Suppl 1):40–8. <u>http://</u> dx.doi.org/10.1080/01926230490425003 PMID:15209402

Maule M, Scélo G, Pastore G, Brennan P, Hemminki K, Tracey E, et al. (2007). Risk of second malignant neoplasms after childhood leukemia and lymphoma: an international study. J Natl Cancer Inst. 99(10):790– 800. <u>http://dx.doi.org/10.1093/jnci/djk180</u> PMID:17505074

Moolgavkar S, Krewski D, Zeise L, Cardis E, Moller H, editors (1999). Quantitative estimation and prediction of human cancer risks. Lyon, France: International Agency for Research on Cancer (IARC Scientific Publications, No. 131).

Pierce DA, Vaeth M (2003). Age-time patterns of cancer to be anticipated from exposure to general mutagens. Biostatistics. 4(2):231–48. http://dx.doi.org/10.1093/biostatistics/4.2.231 PMID:12925519

Preston DL, Pierce DA, Shimizu Y, Cullings HM, Fujita S, Funamoto S, et al. (2004). Effect of recent changes in atomic bomb survivor dosimetry on cancer mortality risk estimates. Radiat Res. 162(4):377–89. <u>http://dx.doi.</u> org/10.1667/RR3232 PMID:15447045

Preston DL, Shimizu Y, Pierce DA, Suyama A, Mabuchi K (2003). Studies of mortality of atomic bomb survivors. Report 13: solid cancer and noncancer disease mortality: 1950–1997. Radiat Res. 160(4):381–407. <u>http://dx.doi.org/10.1667/RR3049 PMID:12968934</u>

Swerdlow AJ, Higgins CD, Smith P, Cunningham D, Hancock BW, Horwich A, et al. (2011). Second cancer risk after chemotherapy for Hodgkin's lymphoma: a collaborative British cohort study. J Clin Oncol. 29(31):4096–104. http://dx.doi.org/10.1200/ JCO.2011.34.8268 PMID:21969511

Travis LB (2006). The epidemiology of second primary cancers. Cancer Epidemiol Biomarkers Prev. 15(11):2020–6. <u>http://dx.doi.org/10.1158/1055-9965.EPI-06-0414</u> PMID:17057028

Travis LB, Curtis RE, Glimelius B, Holowaty EJ, Van Leeuwen FE, Lynch CF, et al. (1995). Bladder and kidney cancer following cyclophosphamide therapy for non-Hodgkin's lymphoma. J Natl Cancer Inst. 87(7):524– 30. <u>http://dx.doi.org/10.1093/jnci/87.7.524</u> PMID:7707439

Travis LB, Rabkin CS, Brown LM, Allan JM, Alter BP, Ambrosone CB, et al. (2006). Cancer survivorship – genetic susceptibility and second primary cancers: research strategies and recommendations. J Natl Cancer Inst. 98(1):15–25. <u>http://dx.doi.org/10.1093/jnci/</u> djj001 PMID:16391368

Vogel EW, Barbin A, Nivard MJ, Stack HF, Waters MD, Lohman PH (1998). Heritable and cancer risks of exposures to anticancer drugs: inter-species comparisons of covalent deoxyribonucleic acid-binding agents. Mutat Res. 400(1–2):509–40. <u>http://dx.doi.</u> org/10.1016/S0027-5107(98)00060-8 PMID:9685708

Vogel EW, Nivard MJ, Ballering LA, Bartsch H, Barbin A, Nair J, et al. (1996). DNA damage and repair in mutagenesis and carcinogenesis: implications of structure-activity relationships for cross-species extrapolation. Mutat Res. 353(1–2):177–218. <u>http://dx.doi.org/10.1016/0027-5107(96)00032-2 PMID:8692191</u>