SECTION OF MOLECULAR PATHOLOGY (MPA)

Section Head Dr Hiroko Ohgaki The Section of Molecular Pathology studies the molecular basis of human neoplasms, in particular brain tumours, using tumour samples from patients with excellent clinical data and follow-up. We correlate histologically recognised phenotypes with genotypes and expression profiles, with the objectives of elucidating the molecular basis and genetic pathways that are operative in human neoplasms; identifying molecular markers for improvement of tumour diagnoses and classification; identifying genetic factors that predict sensitivity to treatment, tumour progression and patient outcome; and using genetic data to identify the etiology of human cancers. Since 2006, the Section of Molecular Pathology has also been responsible for the 4^{th} edition of the World Health Organization (WHO) Classification of Tumours Series (WHO Blue Books). The second volume, WHO Classification of Tumours of the third volume (WHO Classification of Tumours of the Digestive System) is in the editing stages.

In its current configuration the Section consists of a single Group, the Molecular Pathology Group (MPA), with the objectives stated above. A few of its more important projects over the Biennium are detailed below.

MOLECULAR PATHOLOGY GROUP

Head Dr Hiroko Ohgaki

Technical assistance

Mrs Anne-Marie Camus-Randon Miss Christine Carreira (histology laboratory)

Secretariat Mrs Pascale Collard

Database Mr Sébastien Antoni

Visiting scientists and postdoctoral fellows

Dr Shengqing Lu (from November 2007) Dr Young Ho Kim (from May 2009) Dr Takuya Watanabe (March 2007–June 2009) Dr Izabela Zawlik (July 2006-July 2008) Dr Jian Huang (June 2006-December 2008) Dr Sumihito Nobusawa (from April 2008) *IDH1* MUTATIONS AS MOLECULAR SIGNATURE AND PREDICTIVE FACTOR OF SECONDARY GLIOBLASTOMAS AND AS EARLY EVENTS IN THE DEVELOPMENT OF ASTROCYTOMAS AND OLIGODENDROGLIOMAS

IDH1 encodes isocitrate dehydrogenase 1, which participates in the citric acid cycle and was first reported to be mutated in a study of sequencing >20 000 protein coding genes (Parsons et al. Science. 321:1807-1812 2008). We assessed IDH1 mutations in 321 gliomas of various histological types and biological behavior. A total of 130 IDH1 mutations were detected, all located at codon 132; 91% of these were G->A mutations (R132H). IDH1 mutations were frequent in low-grade diffuse astrocytomas (88%) and in secondary glioblastomas that developed through progression from low-grade diffuse or anaplastic astrocytoma (82%). Similarly high frequencies of IDH1 mutations were found in oligodendrogliomas (79%) and oligoastrocytomas (94%). Analysis of multiple biopsies from the same patient (51 cases) showed that there was no case in which an IDH1 mutation occurred after acquisition of a TP53 mutation or loss of 1p/19g, suggesting that IDH1 mutations are very early events in gliomagenesis and may affect a common glial precursor cell population. IDH1 mutations were co-present with TP53 mutations in 63% of low-grade diffuse astrocytomas, and with LOH 1p/19g in 64% of oligodendrogliomas. They were rare in pilocytic astrocytomas (10%) and primary glioblastomas (5%), and absent in ependymomas.

Our analyses of *IDH1* mutations in glioblastomas from a populationbased study (407 cases) showed that approx. 9% of all glioblastomas in a population contain *IDH1* mutations, and that glioblastoma patients with IDH1 mutations are significantly younger (mean 47.9 years) and show longer survival than those without IDH1 mutations. IDH1 mutations were frequent in glioblastomas diagnosed as secondary (22/30; 73%), but rare in primary glioblastomas (14/377; 3.7%: P<0.0001). IDH1 mutations as genetic marker of secondary glioblastoma corresponded to the respective clinical diagnosis in 95% of cases. IDH1 mutations are the therefore most reliable molecular marker of secondary glioblastomas available and should be used to complement clinical criteria to distinguish them from primary glioblastoma. The frequent presence of IDH1 mutations in secondary glioblastomas and their almost complete absence in primary glioblastomas reinforces the concept that despite their histological similarity, these subtypes are genetically and clinically distinct entities.

We assessed *IDH1* mutations in brain tumors diagnosed in patients from 3 families with Li-Fraumeni syndrome. We identified *IDH1* mutations in 5 astrocytomas that developed in carriers of a *TP53* germline mutation. Without exception, all were R132C, which in sporadic astrocytomas accounts for <5% of *IDH1* mutations. This remarkably selective occurrence of R132C mutations may reflect differences in the sequence of genetic events, with a preference for R132C mutations in astrocytes or precursor cells that already carry a germline *TP53* mutation.

Role of mutations in the Nijmegen breakage syndrome gene (NBSI) in brain tumours

Nijmegen breakage syndrome, caused by *NBS1* germline mutations, is a rare autosomal recessive disease with clinical features that include microcephaly, increased radiosensitivity and predisposition to cancer. *NBS1* plays a key role in DNA double-strand break repair and the maintenance of genomic stability. There may be functional interactions between *NBS1* and the *TP53* pathways.

We assessed whether NBS1 mutations play a role in the pathogenesis of sporadic medulloblastomas. Screening for mutations in the NBS1 gene (all 16 exons) and the TP53 gene (exons 5-8) revealed that 7 of 42 (17%) medulloblastomas carried a total of 15 NBS1 mutations (10 missense point mutations and 5 intronic splicing mutations). Of five medulloblastomas with TP53 mutations, four (80%) contained NBS1 mutations, and there was a significant association between TP53 mutations and NBS1 mutations (P=0.001), suggesting that medulloblastomas characterised by NBS1 mutations typically associated with mutational inactivation of the TP53 gene.

We also screened 87 glioblastomas for *NBS1* mutations, and showed 12 *NBS1* mutations) in 9 of 28 (32%) primary (de novo) glioblastomas carrying two or more *TP53* mutations. In contrast, *NBS1* mutations were not detected in 19 primary glioblastomas with one *TP53* mutation, nor in 21 primary glioblastomas without *TP53* mutations. These results suggest that multiple *TP53* mutations in some glioblastomas are due to deficient repair of DNA double-strand breaks caused by mutational inactivation of the *NBS1* gene.

PROMOTER METHYLATION AND POLYMORPHISMS OF THE *MGMT* GENE IN GLIOBLASTOMAS: A POPULATION-BASED STUDY

O⁶ - M e t h y l g u a n i n e - D N A methyltransferase (*MGMT*) is a repair enzyme that removes promutagenic O⁶-methylguanine adducts in DNA in order to protect cells from acquisition of G:C->A:T mutations. MGMT promoter methylation and polymorphisms may affect MGMT expression and activity. We assessed MGMT promoter methylation polymorphisms (Leu84Phe, and Ile143Val, c.-56C>T) in 371 glioblastomas diagnosed at the population level. MGMT methylation was observed in 165 (44%) glioblastomas, with a higher frequency in females than males (53% vs. 39%; P=0.0106), and in secondary than primary glioblastomas (73% vs. 43%, P=0.0074). The frequency of TP53 G: C->A:T mutations in glioblastomas with MGMT methylation was 25%, which was significantly higher than that in glioblastomas without MGMT methylation (16%; P=0.0385). The MGMT 143 Val allele was significantly less frequent in glioblastomas than in a healthy European Caucasian population, and was associated with longer survival than those with the MGMT 143 Ile allele (hazard ratio 0.70; 95% CI=0.48-1.01). These results suggest that MGMT methylation may be associated with susceptibility to acquire TP53 G:C->A:T mutations, and that MGMT polymorphisms may affect the risk and prognosis of glioblastomas.

Common polymorphisms in the MDM2 and TP53 genes and the relationship between TP53 mutations and patient outcomes in glioblastomas

MDM2 SNP309 is associated with a younger age of tumour onset in patients with Li-Fraumeni syndrome, and TP53 codon 72 polymorphism decreases its apoptotic potential. Glioblastomas frequently show genetic alterations in the TP53 pathway. We assessed MDM2 SNP309 in 360 glioblastomas, and correlated these with patient age and survival, as well as other alterations in the TP53 pathway. Frequencies of the MDM2 SNP309 T/T, T/G and G/G genotypes in glioblastomas were 40%, 46% and 14%, respectively. Multivariate analysis showed that the MDM2 SNP309 G/G allele was significantly associated with favourable outcome in female glioblastoma patients (hazard ratio 0.54; 95% CI=0.32-0.92). There was a significant association between MDM2 SNP309 G alleles and TP53 codon 72

Pro/Pro in glioblastomas. Glioblastoma patients with *TP53* codon 72 Pro/Pro genotype were significantly younger than Arg/Arg carriers (mean 50.2 vs. 56.1 years; P=0.018). Multivariate analysis showed that those with *TP53* codon 72 Arg/Pro allele had significantly shorter survival than those with the Arg/Arg allele (hazard ratio 1.35; 95% CI=1.07–1.71). Detailed analyses revealed that the *TP53* codon 72 Pro allele was significantly associated with shorter survival among patients with glioblastomas carrying a *TP53* mutation, and among those treated with surgery plus radiotherapy.

Whole genome Amplification for Array CGH using DNA extracted From formalin-fixed paraffinembedded histological sections

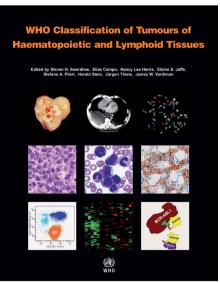
Array comparative genomic hybridization (CGH) is useful to assess genomewide chromosomal imbalance, but the requirement for relatively large amounts of DNA can be a limitation, in particular for samples extracted from small tumour areas on paraffin sections. Whole genome amplification (WGA) can be carried out before array CGH to obtain sufficient DNA, but the possibility of artefacts due to biased amplification cannot be excluded. We optimized the WGA protocol to generate sufficient DNA with minimum amplification bias. Using formalin-fixed paraffin-embedded histological sections of tumours carrying known TP53 mutations, LOH 1p, LOH 10g, LOH 19g, and EGFR amplification, we first optimised the protocol so that these genetic alterations are detected after WGA. We found that a ligation step before WGA is important, as it allows a short reaction time with Phi29 to generate WGA-DNA with greatly decreased amplification bias. Using template >150 ng of DNA, a ligation step before WGA, and a short reaction time with Phi29 DNA polymerase (<1.5 h), we obtained WGA-DNA (>4 g) with minimum amplification bias (<3-fold). Using this protocol, we carried out array CGH (Agilent 105K) before and after WGA. Pearson correlation analysis indicated a significant positive correlation in array CGH results between DNA before and after WGA (P<0.0001). These results suggest that genetic analyses are possible using WGA-DNA extracted from paraffin sections, but that they should be carried out with a carefully optimised and controlled protocol.

WHO CLASSIFICATION OF TUMOURS SERIES (WHO BLUE BOOKS)

The objective of this project is to establish a pathological and genetic classification and grading of human tumours that is accepted and used worldwide. Without clearly defined clinical and histopathological diagnostic criteria and, more recently, genetic and expression profiles, epidemiological studies and clinical trials are difficult to conduct. This project therefore has a substantial impact in not only pathology communities, but also cancer registration, epidemiology studies, clinical trials, and cancer research in general.

IARC has been responsible for this book project since the 3rd edition (2000–2005), which covered all organ sites in 10 volumes. Diagnostic criteria, pathological features and associated genetic alterations were described in a strictly disease-oriented manner. For each volume, 10 000–35 000 copies were printed and distributed worldwide.

The current edition (4th edition) was initiated in 2006, with four new series editors (Dr Fred Bosman, University of Lausanne, Switzerland; Dr Elaine Jaffe, National Institutes of Health, Bethesda, USA; Dr Sunil Lakhani, University of Queensland, Brisbane, Australia; and Dr Hiroko Ohgaki, IARC). The first volume of the 4th edition, *Tumours of the Nervous* System, was published in June 2007. The second volume, Tumours of the Haematopoietic and Lymphoid Tissues, was published in September 2008, and over 30 000 copies have already been printed and distributed worldwide. We are currently preparing the 3rd volume, Tumours of the Digestive System, with 4 volume editors (Dr F. Bosman, Lausanne, Switzerland; Dr F. Carneiro, Porto, Portugal; Dr R.H. Hruban, Baltimore, USA; and Dr N.D. Theise, New York, USA) and with >110 contributors. The consensus and editorial conference is scheduled for December 2009, and the book is scheduled to be published in 2010.



The Section of Molecular Pathology is grateful to the following scientists for their collaboration in its projects:

Dr F. Berger, Grenoble, France Dr F. Bosman, Geneva, Switzerland Dr D.J. Brat, Atlanta, USA Dr E. Campo, Barcelona, Spain Dr F. Carneiro, Porto, Portugal Dr W.K. Cavenee, La Jolla, USA Dr I. Ciernik, Zurich, Switzerland Dr M.A. Grotzer MA, Zurich, Switzerland Dr N.L. Harris, Boston, USA Dr F. Heppner, Zurich, Switzerland Dr E Hewer, Zurich, Switzerland Dr R.H. Hruban, Baltimore, USA Dr J.P. Issartel, Grenoble, France Dr E.S. Jaffe, Bethesda, USA Dr P. Kleihues, Zurich, Switzerland Dr H. Klocker, Innsbruck, Austria Dr S.R. Lakhani, Herston, Australia Dr UM Lutolf, Zurich, Switzerland Dr M. Mittelbronn, Frankfurt, Germany

Dr Y. Nakazato, Gunma, Japan Dr H.-K. Ng, Hong Kong, China Dr J. Pang, Hong Kong, China Dr T. Pietsch, Bonn, Germany Dr S. Pileri, Bologna, Italy Dr N. Probst, Zurich, Switzerland Dr S. Rutkowski, Wurzburg, Germany Dr G. Schafer, Innsbruck, Austria Dr H. Stein, Berlin, Germany Dr S.H. Swerdlow, Pittsburgh, USA Dr N.D. Theise, New York, USA Dr J. Thiele, Cologne, Germany Dr J.W. Vardiman, Chicago, USA Dr A. Vital, Bordeaux, France Dr W.A. Weiss, San Francisco, USA Dr S. Wellek, Mannheim, Germany Dr M. Weller, Zurich, Switzerland Dr H. Yokoo, Gunma, Japan

The financial support from the following bodies is gratefully acknowledged:

Foundation for Promotion of Cancer Research, Japan MEDIC Foundation

PUBLICATIONS

ORIGINAL ARTICLES

Huang J. Grotzer MA, Watanabe T, Hewer E, Pietsch T, Rutkowski S, Ohgaki H. Mutations in the Nijmegen breakage syndrome gene in medulloblastomas. Clin Cancer Res. 14: 4053-4058 (2008)

Yokoo H, Tanaka Y, Nobusawa S, Nakazato Y, Ohgaki H. Immunohistochemical and ultrastructural characterization of brain tumors in S100beta-verbB transgenic rats. Neuropathology, 28: 591-598 (2008)

Zawlik I, Vaccarella S, Kita D, Mittelbronn M, Franceschi S, Ohgaki H. Promoter methylation and polymorphisms of the MGMT gene in glioblastomas: a population-based study. Neuroepidemiology 32: 21-29 (2009)

Zawlik I, Kita D, Vaccarella S, Mittelbronn M, Franceschi, S, Ohgaki H. Common polymorphisms in the MDM2 and TP53 genes and the relationship between TP53 mutations and patient outcomes in glioblastomas. Brain Pathology, 19: 188-194 (2009)

Watanabe T, Nobusawa S, Lu S, Huang J, Mittelbronn M, Ohgaki H. Mutational inactivation of the Nijmegen Breakage Syndrome gene (NBS1) in glioblastomas is associated with multiple TP53 mutations. J. Neuropathol. Exp. Neurol. 68: 210-215 (2009)

Watanabe T, Nobusawa S, Kleihues P, Ohgaki H. IDH1 mutations are early events in the development of astrocytomas and oligodendrogliomas. Am J Pathology 174: 1149-1153 (2009)

Huang J, Pang J, Watanabe T, Ng H-K, Ohgaki H. Whole genome amplification for array comparative genomic hybridization using DNA extracted from formalin-fixed paraffin-embedded histological sections. Journal of Molecular Diagnostics 11: 109-116 (2009)

Kita D, Ciernik I, Vaccarella S, Franceschi S, Kleihues P, Lutolf UM, Ohgaki H. Age as predictive factor in glioblastomas: population-based study. Neuroepidemiology 33: 17-22 (2009)

Sun X, Huang J, Homma T, Kita D, Klocker H, Schafer G, Boyle P, Ohgaki H. Genetic alterations in the PI3K pathway in prostate cancer. Anticancer Res 29: 1739-1743 (2009)

Watanabe T, Vital A, Nobusawa S, Kleihues P, Ohgaki H. Selective acquisition of IDH1 R132C mutations in astrocytomas associated with Li-Fraumeni syndrome. Acta Neuropathol 117: 653-656 (2009)

Nobusawa S, Watanabe T, Kleihues P, Ohgaki H. IDH1 mutations as molecular signature and predictive factor of secondary glioblastomas. Clinical Cancer Res in press

BOOK CHAPTERS AND REVIEWS

Ohgaki H. Brain Cancer. In: Prognosis in Advanced Cancer. Glare P, Christakis NA (eds.) Oxford University Press pp. 201-213 (2008)

Ohgaki H. Epidemiology of brain tumors. Methods Mol. Biol. 472: 323-342 (2009)

Ohgaki H. and Kleihues P. Genetic alterations and signaling pathways in the evolution of gliomas. Cancer Science, submitted.