

CHAPTER 7. POLYCYCLIC AROMATIC HYDROCARBONS IN AMBIENT AIR AND CANCER

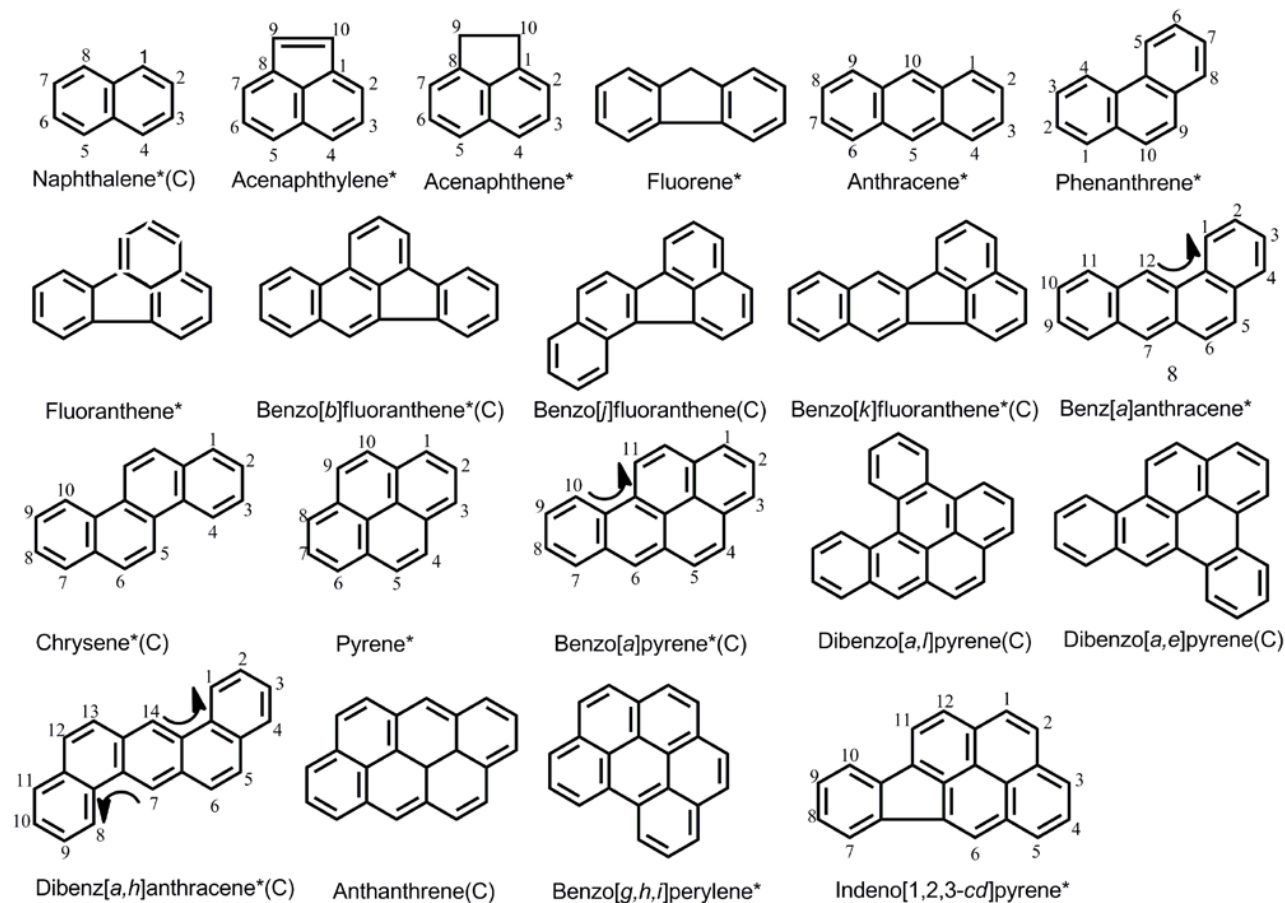
Erik Dybing, Per E. Schwarze, Per Nafstad, Katarina Victorin, and Trevor M. Penning

Polycyclic aromatic hydrocarbons (PAHs), which are generated from the incomplete combustion of organic (carbonaceous) material, are ubiquitous contaminants in ambient air ([IARC, 1983](#), 1984a, 1984b, [1985](#), [2010](#); [WHO, 1998](#)). Their occurrence in the air we breathe has been substantial during the past centuries due to emissions from industrial processes and energy production, motor vehicular traffic, incineration of refuse, and residential heating.

PAHs consist of two or more fused aromatic rings made up of carbon and hydrogen atoms. The ring systems can be present in multiple configurations and may be unsubstituted or substituted. PAHs range from semivolatile molecules to molecules with high boiling points. Thus, they may be found both in the gas and the particulate phase of ambient air or in mixtures of both phases. About 500 different PAHs have been detected in air, but often the measurements focus on benzo[*a*]pyrene (B[*a*]P) as a representative of the whole PAH family ([WHO, 1998](#); [Boström et al., 2002](#)). Many of the PAHs in ambient air are carcinogenic ([IARC, 1983](#), 1984a, 1984b, [1985](#), [2010](#)) ([Figure 7.1](#)), and a recent reassessment of their carcinogenic potential led to B[*a*]P being

upgraded to a Group 1 *known human carcinogen* ([IARC, 2010](#)). Thus there is considerable concern about the relationship between PAH exposure in the ambient air and the potential to contribute to human cancer incidence. The United States Environmental Protection Agency (EPA) monitors 16 priority PAHs in air due to health concerns: naphthalene, acenaphthylene, acenaphthene, fluorene, anthracene, phenanthrene, fluoranthene, pyrene, chrysene, benz[*a*]anthracene, benzo[*b*]fluoranthene, benzo[*k*]fluoranthene, B[*a*]P, indeno[1,2,3-*cd*]pyrene, benzo[*g,h,i*]-perylene, and dibenz[*a,h*]anthracene (in order of number of aromatic rings per structure) ([Figure 7.1](#)). Of particular note is that several PAHs (naphthalene, chrysene, benzo[*b*]fluoranthene, benzo[*k*]fluoranthene, B[*a*]P, dibenz[*a,h*]anthracene, dibenzo[*a,e*]pyrene and dibenzo[*a,l*]pyrene, and anthanthrene) have been found to be carcinogenic in experimental animals after inhalation or intratracheal ingestion, increasing concern about the levels of these carcinogens in ambient air ([Figure 7.1](#)).

Fig 7.1 PAHs in ambient air.



An asterisk denotes a United States Environmental Protection Agency priority pollutant. (C) indicates that the compound is carcinogenic by inhalation or intratracheal administration in experimental animals. Source: [Park and Penning \(2008\)](#); reproduced with permission from John Wiley & Sons.

PAH emissions in ambient air

A recent global atmospheric emission inventory of PAHs ([Zhang and Tao, 2009](#)) showed that the emission from the 16 priority PAHs listed by the EPA was 520 000 tonnes per year. Anthropogenic sources of total PAHs in ambient air emissions are greater than those that come from natural events such as forest fires and volcanic eruptions.

Apart from localized risk at or near the source of emission, PAHs can be dispersed regionally and intercontinentally through atmospheric long-range transport. For example, PAHs

emitted from East Asia are transported to the west coast of the USA, and PAHs emitted in the Russian Federation influence atmospheric PAH concentrations in the Arctic ([Zhang and Tao, 2009](#)). The annual PAH emission from Asian countries is 290 000 tonnes (55% of the total); the amounts from China (114 000 tonnes per year) and India (90 000 tonnes per year) are the major contributors. The USA is the third largest emitter of PAHs, at 32 000 tonnes per year. By contrast, European countries account for only 9.5% of the total PAH emissions annually ([Zhang and Tao, 2009](#)). The contribution of the various anthropogenic sources of PAHs to the total emission

profile can vary by country and region. The global sources of PAH emissions are shown in [Table 7.1](#), and the main sources of PAHs in six European countries are shown in [Table 7.2](#).

The largest emission of PAHs globally comes from incomplete combustion of organic material, and the largest single source is from the combustion of biofuels. Biofuel is a single type of primary solid biomass (e.g. animal dung or peat) ([Zhang and Tao, 2009](#)). Burning biomass fuels such as wood on indoor open-pit stoves is common in developing areas, leading to harmful exposures to particulate matter < 2.5 µm in diameter (PM_{2.5}), carbon monoxide (CO), and PAHs, which can be significantly reduced by the introduction of modern stoves ([Li et al., 2011](#)). Anthropogenic sources include PAHs that come from incomplete combustion processes (especially biofuels) and those that are made commercially, are by-products of industrial processes, or are generated from vehicle emissions, cooking, food preservation, and first- and second-hand cigarette smoke.

Anthropogenic sources of PAHs in ambient air

Commercial production

PAHs produced commercially include naphthalene, acenaphthene, phenanthrene, fluoranthene, and pyrene; however, only naphthalene is used directly without further processing, as a moth repellent.

Industrial processes

Many PAHs are released into the atmosphere during industrial processes such as coal coking and petroleum refining. It is estimated that coal coking was responsible for the release of thousands of tonnes of PAHs per year in different countries during the 1980s and early 1990s. Reduced coke production and technical

improvements have led to reductions in PAH emissions from this source. Little is known about the composition of these PAH emissions ([WHO, 1998](#)). In petroleum refining, most of the emissions consist of smaller two- and three-ring compounds (94–99%, depending on the process studied) ([IARC, 1989](#)). Thus, the composition of PAHs from combustion (pyrogenic) versus the composition of PAHs from petroleum refining (petrogenic) can be widely different. Other industrial sources with significant PAH emissions are carbon black plants, wood preservation (creosote) plants, the asphalt and bitumen industry, aluminium production (Söderberg electrodes), iron and steel production, foundries, tyre production, power plants, waste incinerators, and stubble burning ([WHO, 1998](#)). Further restrictions may lead to lower PAH emissions from these industries ([CORINAIR, 1997](#)).

Estimation of the PAH emissions for six European countries indicates that the industrial sources contribute PAHs in the same range as mobile sources ([Table 7.2](#); data from [CORINAIR, 1997](#)).

Residential sources

Domestic heating with oil and wood stoves leads to considerable PAH emissions in northern European countries, and especially in Scandinavia ([Boström et al., 2002](#)). In Sweden, the emissions from wood-fired domestic heating are estimated to be about 100 tonnes per year, with minor contributions from oil combustion. Environmental tobacco smoke is also a considerable source of indoor air pollution and contamination within the home ([Hoh et al., 2012](#)).

Motor vehicle emissions

The amount of PAHs released into the air from vehicles has been reduced considerably by the introduction of three-way converters. However, older diesel and gasoline cars with a

Table 7.1 Main sources of emission for the United States Environmental Protection Agency 16 priority PAHs in China, India, and the USA

Source	Global	China	India	USA
Biofuel	56.7%	66.4%	92.5%	9.1%
Wild fire	17.0%	0%	0%	3.3%
Consumer product use	6.9%	0.9%	0.6%	35.1%
Traffic oil	4.8%	2.0%	IS	23.0%
Domestic coal	3.7%	10.7%	1.3%	IS
Coke production	3.6%	14.4%	IS	IS
Petroleum refining	2.4%	1.0%	IS	8.7%
Waste incineration	1.9%	IS	IS	9.5%
Aluminium electrolysis	1.4%	IS	IS	1.9%
Open straw burning	IS	2.0%	3.2%	IS
Gasoline distribution	IS	IS	IS	3.0%
Aerospace industry	IS	IS	IS	2.5%
Other	1.5%		2.7%	3.9%
Tonnes in thousands	530	114	90	32

IS, insignificant.

Compiled from [Zhang and Tao \(2009\)](#).

catalytic converter of outmoded design have 5–10 times higher PAH emissions than modern cars. In addition, cold start at temperatures below the standardized cold start (23 °C), and especially at temperatures below 0 °C, results in a several-fold increase in PAH emissions. Several other technical variations lead to varying emissions, for example spark ignition engines ([WHO, 1998](#)). The total amounts of PAHs emitted from vehicles vary between countries; in the USA this can be as high as 6000 tonnes per year, and in six European countries the amount is about 400 tonnes per year ([Table 7.1](#) and [Table 7.2](#)).

As might be expected, not all PAHs contribute equally to the emissions into ambient air. [Table 7.3](#) lists a typical PAH profile in ambient air arising from different sources.

Human exposure

PAHs may be found in the gas and particulate phases (see Chapter 1). The levels given below frequently reflect the levels of discrete PAHs in the particulate phase and are often given as the sum of a limited number of PAH components.

B[a]P is the traditional marker for PAH exposure. Several additional PAH components have been proposed as emission markers, for example fluoranthene, B[a]P, and benzo[b]fluoranthene. [Boström et al. \(2002\)](#) suggested the use of the following set of PAHs as emission and effect markers for monitoring air pollution: B[a]P, fluoranthene, phenanthrene, methylanthracenes/phenanthrenes, pyrene, benzo[b]fluoranthene, benzo[k]fluoranthene, indeno[1,2,3-*cd*]pyrene, benzo[*g,h,i*]-perylene, dibenz[*a*]anthracene, and dibenzo[*a,l*]pyrene. This list is quite similar to the 16 priority PAHs listed by the EPA ([Figure 7.1](#)). In some studies, the total PAH exposure is given as B[a]P toxic equivalency concentrations. In this approach, individual components are measured and ranked relative to B[a]P in terms of carcinogenicity. For example, chrysene has 1/1000th of the carcinogenicity of B[a]P and has a toxic equivalency concentration of 0.001. These calculations are used to estimate human health risk and can be used to calculate incremental lifetime cancer risk (ILCR). $ILCR = \text{exposure } (\mu\text{g}/\text{kg}/\text{day}) \times \text{cancer slope factor } (\mu\text{g}/\text{kg}/\text{day})$. The ILCR is considered negligible when it is less than 1 in 10^5 .

Table 7.2 Main source sectors for PAHs in 1994 in six European countries (Austria, Denmark, Germany, Luxembourg, Norway, and the United Kingdom)

Sector	PAH emissions	
	Amount (tonnes per year)	Percentage of total
Combustion of energy and transformation industries	6.1	0.3
Non-industrial combustion plants plus wood burning	1120	60
Combustion in manufacturing industry	63	3.4
Production processes	248	13
Road transport	383	20
Other mobile sources	10	0.5
Waste incineration	30	1.6
Agriculture and forestry	1	< 0.1
Natural sources	8	0.4
Total (approximately)	1900	

Reproduced from [Boström et al. \(2002\)](#).

(less than 1 additional cancer case per 100 000 persons), and the cancer slope factor is based on the extrapolation of a dose–response curve for tumorigenicity seen at high dose in experimental animals.

Background levels of PAHs in remote locations have been measured between 0.01 ng/m³ and 0.1 ng/m³ for individual PAH components ([WHO, 1998](#)). In rural districts the levels were approximately 10 times higher, whereas in city streets levels may amount to 50 ng/m³ or more of the more abundant individual PAHs ([Boström et al., 2002](#)). Total PAHs in the centre of Stockholm, Sweden, ranged from below 100 ng/m³ to 200 ng/m³. The most abundant PAH was phenanthrene. In other cities higher levels of individual PAHs have been measured ([WHO, 1998](#); [Binková et al., 2003](#)). PAH was measured in the gas and particulate phase over summer and winter sampling periods in Kocaeli, Turkey. Σ_{13} PAH in the gas and particulate phases ranged from 6.2 ng/m³ dibenz[*a,h*]anthracene to 98.6 ng/m³ phenanthrene in the winter, and from 3.0 ng/m³ benz[*a*]anthracene to 35.1 ng/m³ phenanthrene in the summer. The most abundant PAH in both sampling periods was phenanthrene, followed by fluoranthene and pyrene. B[*a*]P toxic

equivalency concentrations were found to be 3-fold higher in the winter months ([Gaga et al., 2012](#)). A similar outcome was observed in a study of children aged 5–6 years (n = 260) in New York City when measurements were conducted in the heating and non-heating seasons ([Jung et al., 2010](#)). In the United Kingdom, the Toxic Organic Micropollutants programme measured temporal trends in PAH in the atmosphere from 1991 to 2005 at six different sampling sites. Most showed a reduction in PAH levels and had concentrations that were lower than the new air quality standard of 0.25 ng/m³. However, this value was exceeded in urban areas in the winter months ([Meijer et al., 2008](#)).

Indoor PAH levels usually range from 1 ng/m³ to 50 ng/m³ due to tobacco smoke and residential heating with wood, coal, and other materials ([WHO, 1998](#)). Environmental tobacco smoke is a major contributor to air pollution and dust, and surfaces remain contaminated long after the smoking has ceased (called third-hand smoke). Measurement of PAHs in settled household dust in 132 homes showed that total PAHs were 990 ng/g in smoking households versus 756 ng/g in nonsmoking households, and when corrected

Table 7.3 Mean profiles of individual PAHs in ambient air (relative to benzo[*a*]pyrene = 1.0)

Compound	Point source	Near mobile source	Home heating	Transport	Geometric mean
Anthracene	5.5	7.6	1.0	1.8	2.9
Phenanthrene	38	200	39	43	60
Fluoranthene	14	48	12	13	18
Pyrene	9.3	28	11	7.1	12
Benz[<i>a</i>]anthracene	1.4	0.82	1.0	0.78	0.97
Perylene	0.33	0.25	0.22	0.24	0.26
Benzo[<i>e</i>]pyrene	1.5	1.3	1.6	1.4	1.4
Benzo[<i>g,h,i</i>]perylene	1.4	1.5	2.4	1.3	1.6
Indeno[1,2,3- <i>cd</i>]pyrene	1.5	1.3	1.5	1.4	1.4
Anthanthrene	0.19	0.15	0.13	0.20	0.17
Chrysene and triphenylene	3.0	2.7	3.5	2.9	3.0
Benzofluoranthene	3.6	2.9	3.6	4.4	3.6

Source: [WHO \(1998\)](#); reproduced with permission from the publisher.

for loading (dust/m³), the fold change was greater than 2-fold ([Hoh *et al.*, 2012](#)).

PAHs in the ambient air can react with nitrates, hydroxyl radicals, or ozone, leading to the production of more water-soluble compounds. These compounds are rarely included in routine PAH measurements. However, nitro-PAHs have been detected on soot, and the formation of B[*a*]P-nitroquinone has been identified ([Schauer *et al.*, 2004](#)). Exposure levels of nine different nitroarenes resulting from diesel and gasoline exhaust have recently been reviewed by the International Agency for Research on Cancer; diesel exhaust was ranked as a Group 1 known human carcinogen ([Benbrahim-Tallaa *et al.*, 2012](#)).

Generally the mobile sources differ in their PAH profile, with the heavy diesel vehicles being characterized by lower-molecular-weight components than gasoline vehicles. However, per driven kilometre, total emissions from a gasoline-fuelled car are much lower than emissions from a diesel car. The three-way converter does not change the PAH profile of a gasoline-fuelled car significantly but reduces the total levels considerably. PAH levels vary with season, with higher levels being observed in the winter than in the summer. Data from Stockholm, Sweden, indicate that during

the winter the levels of low-molecular-weight PAHs are increased compared with the summer ([Prevedouros *et al.*, 2004](#)).

Biomonitoring

Significant progress has been made in biomonitoring of human exposure to PAH. External dose can be measured using personalized air monitoring devices where PM is trapped on filters and then analysed for PAH content. Internal dose can be assessed by measuring blood and urinary biomarkers of exposure. Different analytes have been used as biomarkers of PAH exposure and effect. These include measuring PAH metabolites in the urine and intermediate biomarkers of effect (e.g. DNA and haemoglobin adducts). Analysis using urinary metabolites has given the most clear-cut results. Particulate pyrene is well correlated with total PAH in the breathing zone.

Urinary 1-hydroxypyrene may also reflect inter-individual variation in PAH metabolism. Occupational exposure has been found to lead to a 10–100 times greater urinary 1-hydroxypyrene content. Danish bus drivers excreted more 1-hydroxypyrene than mail carriers did, but outdoor working mail carriers had more

PAH metabolites in their urine than those working indoors, indicating the impact of outdoor air pollution ([Hansen et al., 2004](#)). The use of 1-hydroxypyrene as a biomarker of PAH exposure has been criticized on the grounds that pyrene is not a carcinogenic PAH. This has led to the substitution of 3-hydroxy-B[a]P, but sensitive methods of detection have been a challenge. The detection of 3-hydroxy-B[a]P has also been criticized as a biomarker since this metabolite is not derived from any of the known pathways of B[a]P activation.

Measurements of urinary 1-hydroxypyrene-glucuronide, 2-naphthol, and malondialdehyde by synchronous fluorescence spectroscopy or high-performance liquid chromatography were used to evaluate seasonal and regional variations in PAH exposure and oxidative stress in Korean adults and women. Higher levels were found in individuals from industrialized areas and in the winter. Further elevation of 1-hydroxypyrene-glucuronide was observed in children exposed to environmental tobacco smoke ([Yoon et al., 2012](#)). In a study in Chinese children from polluted and non-polluted areas, the levels of nine urinary monohydroxylated PAH metabolites and 8-oxo-2'-deoxyguanosine (8-oxo-dG) were compared. Children from the polluted area had a higher PAH burden than those from the non-polluted area, but no significant difference in 8-oxo-dG levels was noted ([Fan et al., 2012](#)). The effect of involuntary tobacco smoke exposure on urinary levels of 23 monohydroxylated metabolites of PAH in 5060 subjects aged > 6 years was studied in the National Health and Nutrition Examination Survey (NHANES). After correcting for other confounders, significant increases in urinary 1-hydroxypyrene, 2-hydroxyfluorene, 3-hydroxyfluorene, 9-hydroxyfluorene, 1-hydroxypyrene, and 1-2-hydroxy-phenanthrene were observed. Increases of 1.1–1.4-fold for involuntary exposure were noted, which increased to 1.6–6.9-fold

increases when children were actively exposed ([Suwan-ampai et al., 2009](#)).

As there is compelling evidence for the conversion of PAH to diol-epoxides as an activation pathway (see below), there have been recent advances in measuring their corresponding tetraol hydrolysis products in humans. Progress has been made in developing stable isotope dilution liquid chromatographic mass spectrometric methods to detect phenanthrene tetraols ([Hecht et al., 2010](#); [Zhong et al., 2011](#)). Phenanthrene contains a bay region and undergoes similar metabolic transformation to B[a]P to form diol-epoxides, which hydrolyse to tetraols. The detection of phenanthrene tetraols has also been criticized, since it is not a carcinogenic PAH. Recently, methods have been developed to measure urinary B[a]P tetraols with femtomole sensitivity ([Hecht et al., 2010](#)), and these techniques can now be applied to biomonitoring studies.

Efforts have also been made to detect stable covalent diol-epoxide DNA and haemoglobin adducts in exposed humans. Repaired diol-epoxide DNA adducts in blood can be measured using ELISA and chemiluminescence-based methods, while unrepaired DNA adducts can be measured in lymphocytes by [³²P]-postlabelling methods. For example, (+)-7 β ,8 α -dihydroxy-9 α ,10 α -oxo-7,8,9,10-tetrahydro-B[a]P-N²-deoxyguanosine [(+)-anti-B[a]PDE-N²-dGuo] adducts have also been detected in human maternal and umbilical white blood cells after exposure to air pollution, using ELISA-based methods ([Whyatt et al., 1998](#); [Santella, 1999](#)). Total DNA and B[a]P-like DNA adducts were measured by [³²P]-postlabelling in lymphocytes of nonsmoking policemen in Prague (n = 109) working 8 hour shifts. While there was no significant change in total DNA adducts, there was a marked increase in B[a]P-like DNA adducts correlated to personal exposure to PAHs collected on respirable particles ([Topinka et al., 2007](#)). Diol-epoxide DNA adducts are

short-lived; therefore, attention has also focused on the development of methods to detect haemoglobin diol-epoxide adducts since the half-life of the red blood cell is 7–10 days ([Day et al., 1990](#)).

Toxicokinetics, including metabolic activation

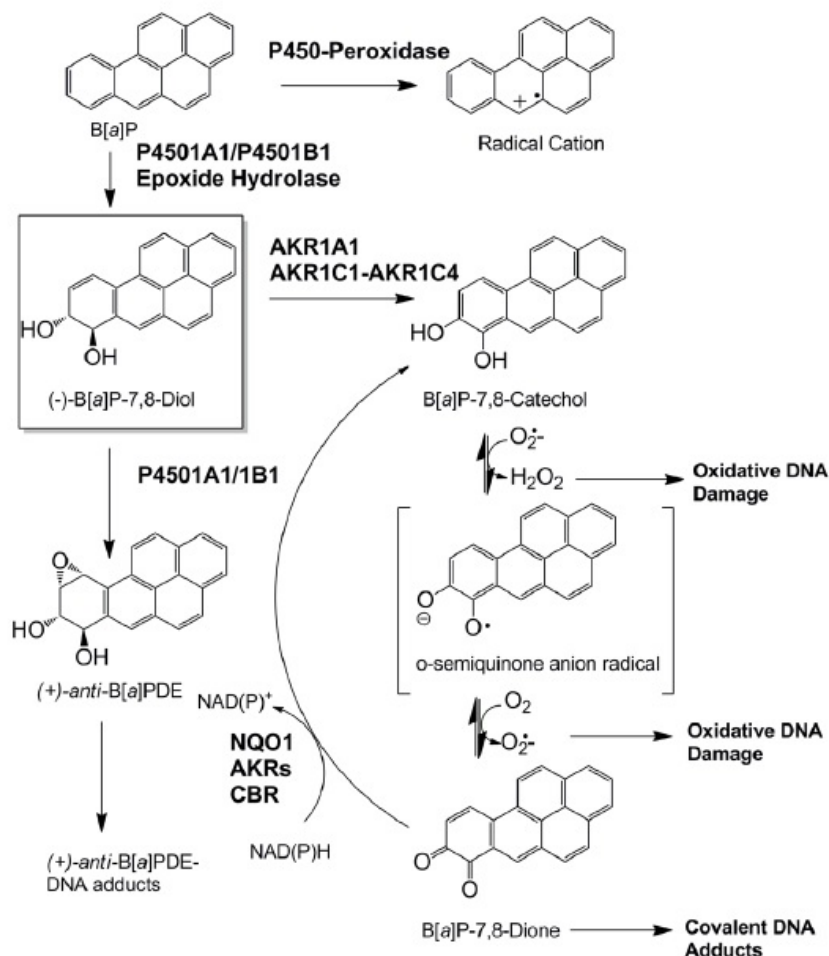
Parent PAHs have low chemical reactivity and must be metabolically activated to electrophilic intermediates to exert their carcinogenic effects ([Sims and Grover, 1974](#); [Conney 1982](#); [Thakker et al., 1985](#)). Three pathways of PAH activation have been proposed in the literature and are best exemplified with B[a]P ([Figure 7.2](#)). In the first pathway, B[a]P is metabolically activated by either P450 peroxidase or another peroxidase by acting as a co-reductant of complex-1 (Fe^{V}). This leads to a radical cation on the most electron-deficient C6 atom, which is highly reactive and capable of forming unstable C8-guanine [8-(benzo[a]pyren-6-yl)guanine], N7-guanine [7-benzo[a]pyren-6-yl]guanine], and N7-adenine [7-benzo[a]pyren-6-yl]adenine] depurinating DNA adducts ([Cavalieri and Rogan, 1995](#)). Evidence for this pathway comes from *in vitro* reactions with B[a]P, microsomes, and a peroxide substrate, which has led to the trapping of DNA adducts, as well as from mouse skin studies ([Cavalieri et al., 1990, 1991](#)). Data exist that B[a]P and dibenzo[a,l]pyrene can exert their tumorigenicity through this mechanism in mouse skin and rat mammary gland ([Cavalieri et al., 1991, 2005](#)) In addition, trace amounts of B[a]P-depurinating DNA adducts have been detected in the urine of smokers and in women exposed to household smoke ([Casale et al., 2001](#)). However, apart from this single study, the evidence to support this mechanism due to inhalation exposure to PAH is not strong.

In the second pathway, B[a]P is metabolically activated to vicinal diol-epoxides ([Jerina et al., 1991](#)) formed through a three-step process

involving oxidation and hydrolysis reactions ([Figure 7.2](#)). In the first step, B[a]P is converted preferentially in the lung by the cytochrome P450 isozyme P4501B1 to the major (+)-7R,8S-epoxide and minor (-)-7S,8R-epoxide. In the second step, the 7R,8R-*trans*-dihydrodiol is predominately formed by the action of epoxide hydrolase. In the third step, diol-epoxide diastereomers are generated by another oxidation reaction via various P450 enzymes, including P4501B1 ([Thakker et al., 1985](#); [Petruska et al., 1992](#); [Guengerich, 1993](#); [Constantin et al., 1994](#); [Cavalieri and Rogan, 1995](#); [Shimada et al., 1999, 2001](#)).

Diol-epoxides have been studied in various animal carcinogenicity models. It has been revealed that the diol-epoxides with the highest carcinogenic activity are in general the *anti*-diastereomers and especially the enantiomers with *R*-absolute configuration at the benzylic arene carbon ([Thakker et al., 1985](#); [Glatt et al., 1991](#)). In studies of interactions of diol-epoxides with DNA, they demonstrate a high preference for the exocyclic amino group of deoxyguanosine and deoxyadenosine, where the major adduct derived from B[a]P is (+)-*anti*-B[a]PDE-N²-dGuo ([Jeffrey, 1985](#); [Gräslund and Jernström, 1989](#); [Jerina et al., 1991](#); [Geacintov et al., 1997](#)). This pathway of metabolic activation has been observed for many PAHs in ambient air, including 5-methylchrysene ([Melikian et al., 1983](#), [Koehl et al., 1996](#)), benz[a]anthracene ([Cooper et al., 1980](#)), benzo[b]fluoranthene ([Ross et al., 1992](#)), B[a]P (as outlined above), dibenz[a,h]anthracene ([Platt et al., 1990](#)), and dibenzo[a,l]pyrene ([Luch et al., 1997, 1999](#)), in *in vitro* systems (cell extracts, microsomes, and cell culture systems), and in some cases in *in vivo* studies in animals and humans. For example, PAHs within airborne PM_{2.5} produced DNA bulky stable adducts in human lung cell co-cultures ([Abbas et al., 2013](#)).

In the third pathway, PAHs are metabolically activated to *o*-quinones by the action of aldo-keto reductases (AKRs) ([Penning et al., 1999](#); [Penning, 2004](#)). For B[a]P, the sequence involves

Fig 7.2 Pathways of PAH activation using benzo[*a*]pyrene as an example.

Source: [Park and Penning \(2008\)](#); reproduced with permission from John Wiley & Sons.

the NAD(P)⁺-dependent oxidation of the 7*R*,8*R*-*trans*-dihydrodiol to a ketol catalysed by AKR1A1, AKR1C1–AKR1C4 ([Figure 7.2](#)). The ketol then spontaneously rearranges to a catechol, which undergoes air-oxidation to yield B[a]P-7,8-dione and reactive oxygen species (ROS) ([Palackal et al., 2001, 2002](#); [Penning et al., 1996](#)). B[a]P-7,8-dione is both electrophilic (will react with DNA) and redox-active. In the presence of reducing equivalents and NQO1, AKRs themselves, and carbonyl reductase, the quinones can be reduced back to the corresponding catechols, and if they are not intercepted a futile redox cycle will ensue in

which NADPH is depleted and ROS is amplified ([Shultz et al., 2011](#)). This pathway of metabolic activation has been observed for several PAHs in ambient air, including phenanthrene, chrysene, 5-methyl-chrysene, benz[*a*]anthracene, and B[a]P in *in vitro* systems (recombinant enzymes) and cultures of human lung cells ([Palackal et al., 2001, 2002](#); [Park et al., 2008b](#)).

Efforts have been made to assess the contribution of each of these pathways to the metabolic activation of B[a]P in human lung cells. Using a stable isotope dilution liquid chromatographic mass spectrometric method, signature

metabolites of each of the three pathways were measured: B[a]P-1,6-dione and B[a]-3,6-dione (radical cation metabolites), B[a]P-tetraol-1 (diol-epoxide metabolites), and B[a]P-7,8-dione (*o*-quinone metabolites) in human bronchoepithelial (H358) cells in the presence and absence of the aryl hydrocarbon receptor (AhR) agonist TCDD. It was found that each of the pathways contributed equally to B[a]P metabolism in the presence and absence of TCDD (Lu *et al.*, 2011).

The rate of absorption of PAHs from the tracheobronchial epithelium after inhalation exposure is determined by their high lipophilicity (Gerde *et al.*, 1993). For lipophilic carcinogens such as B[a]P, the delayed absorption in the airway mucosa is a result of slow passage through the airway epithelium, yielding a very high dose to these target cells. Because of the long retention time, the metabolic activation can be considerable even at low enzyme activities (Bond *et al.*, 1988).

Modes of action

Carcinogenic PAHs are generally positive in short-term tests for mutagenicity (Table 7.4), for example the bacterial *Salmonella* mutagenicity (Ames) assay and the *HPRT*-mammalian cell mutagenicity assay, provided a metabolic activation system is present (Malaveille *et al.*, 1977; MacLeod *et al.*, 1988; Chen *et al.*, 1990; Wei *et al.*, 1993). In the Ames assay, a rat liver S9 activation system is used; in the *HPRT* assay, recombinant P4501A1 and P4501B1 are co-expressed. The mutagenic species has been identified by comparing the mutagenic potency of different PAH metabolites, which demonstrates that of the known metabolites the diol-epoxides are the most potent mutagens (Malaveille *et al.*, 1977). Treatment of a plasmid containing *K-Ras* with the (+)-*anti*-B[a]PDE followed by transfection into NIH3T3 cells led to cell transformation with increased foci in soft agar. Rescue of the plasmid showed that there were single point mutations of

the 12th and 61st codons, which could explain the transformation potential of the diol-epoxide. The dominant mutation observed was a G → T transversion, consistent with DNA-adduct formation on deoxyguanosine (Marshall *et al.*, 1984). One of the most compelling pieces of data has shown that by using ligation-mediated polymerase chain reaction, the (+)-*anti*-B[a]PDE preferentially forms DNA adducts in *hot spots* on the *p53* tumour suppressor gene, which is one of the most mutated genes in human lung cancer. These hot spots correspond to the same codons that are mutated in tumours obtained from humans with lung cancer. The dominant mutation observed was again a G → T transversion, consistent with DNA adduct formation on deoxyguanosine (Denissenko *et al.*, 1996; Hainaut and Pfeifer, 2001).

In a separate *in vitro* study, the mutagenic potency of (±)-*anti*-B[a]PDE and B[a]P-7,8-dione (AKR product) were compared in a yeast-reporter gene assay for *p53* mutation. It was found that B[a]P-7,8-dione was 80-fold more mutagenic than the diol-epoxide provided it was permitted to redox cycle (Yu *et al.*, 2002). In these experiments there was a linear correlation between (±)-*anti*-B[a]PDE mutagenicity and the formation of (+)-*anti*-B[a]PDE-N²-dGuo adducts, and a linear correlation between B[a]P-7,8-dione mutagenicity and the formation of 8-oxo-dGuo adducts (Park *et al.*, 2008a). In addition, B[a]P-7,8-dione gave predominately G → T transversions, consistent with the base mispairing of 8-oxo-dGuo with adenine. The position of the point mutations within *p53* was quite random until there was biological selection for dominance, and then the spectrum of mutations was similar to that seen in lung cancer (Park *et al.*, 2008b). These data suggest that B[a]P-7,8-dione formed by AKRs has the potential to contribute to the carcinogenic mode of action of B[a]P.

Planar PAHs can induce their own metabolism. Compounds such as B[a]P can bind to the AhR (Nebert and Jensen, 1979; Nebert *et al.*,

Table 7.4 Genotoxicity of individual PAHs that are carcinogenic in experimental animals after inhalation or intratracheal instillation

Compound	Results
Anthanthrene	Positive, limited database
Benzo[<i>b</i>]fluoranthene	Positive
Benzo[<i>j</i>]fluoranthene	Positive
Benzo[<i>k</i>]fluoranthene	Positive
Benzo[<i>a</i>]pyrene	Positive
Chrysene	Positive
Dibenz[<i>a,h</i>]anthracene	Positive
Dibenzo[<i>a,i</i>]pyrene	Positive
Indeno[1,2,3- <i>cd</i>]pyrene	Positive
Naphthalene	Negative for gene mutations, positive for clastogenicity <i>in vitro</i>

Source: [WHO \(1998\)](#); reproduced with permission from the publisher.

1993, 2004). This leads to nuclear localization of the liganded AhR, where it can act as a transcription factor by binding to the xenobiotic response element to induce the *CYP1A1* and *CYP1B1* genes ([Denison et al., 1988a, 1988b, 1989](#)), which will result in enhanced monooxygenation of the parent PAH. PAH metabolism leads to the production of electrophiles (e.g. quinones), which can activate the Nrf2-Keap 1 system. Nrf2 acts as a transcription factor and binds to the antioxidant response element to induce γ GCS, *NQO1* and *AKR1C1*–*AKR1C3*, and *AKR1B10* ([Burczynski et al., 1999](#); [Jin and Penning 2007](#); [Penning and Drury, 2007](#)). Importantly, *AKR1C1*–*AKR1C3* are involved in the metabolic activation of PAH *trans*-dihydrodiols to the electrophilic and redox active PAH *o*-quinones, which could further exacerbate PAH activation via induction of AKRs. The PAH *o*-quinones produced by this pathway are also ligands for the AhR ([Burczynski and Penning, 2000](#)). Thus, both the parent PAH and their downstream metabolites can lead to the metabolic activation of PAHs in ambient air.

PAHs may, in addition to initiating carcinogenesis via a genotoxic mechanism, exert promotional effects through various modes of action. Certain PAHs induce inflammatory processes ([Casale et al., 1997](#)). The binding of PAHs to the AhR also leads to transcriptional upregulation of

genes involved in growth as well as biotransformation and differentiation ([Nebert et al., 1993](#)). Studies also indicate the ability of both PAHs and their metabolites to activate kinases involved in survival signalling, thus giving DNA-damaged cells a survival advantage ([Burdick et al., 2003](#)). At higher concentrations some PAHs induce apoptosis ([Solhaug et al., 2004](#)). In addition, PAHs show inhibitory effects on gap junctional intercellular communication ([Upham et al., 1996](#); [Weis et al., 1998](#)).

Carcinogenicity studies in animals

Most investigations of PAH carcinogenesis by the respiratory route are intratracheal instillation studies ([WHO, 1998](#)). In all, 10 PAHs have been found to be carcinogenic in experimental animals after inhalation or intratracheal instillation ([WHO, 1998](#); [NTP, 2000](#)) ([Table 7.5](#)). Only B[*a*]P and naphthalene have been studied by the inhalation route. In one inhalation study in hamsters, groups of 24 males were exposed to B[*a*]P condensed onto sodium chloride particles at concentrations of 2.2, 9.5, and 46.5 mg/m³ for 4.5 hours per day, 7 days per week for the first 10 weeks, then for 3 hours per day for 2 years. Exposure was by nose breathing only. There were no tumours in the controls or in the low-exposure

group. In the other two groups, exposure-related tumours were found in the nasal cavity, larynx, trachea, pharynx, oesophagus, and forestomach, but not in the lung ([Thyssen et al., 1981](#)). [RIVM \(1989\)](#) cites two other inhalation studies with B[a]P not found in the open literature: one in mice ([Knizhnikow et al., 1982](#); see [RIVM, 1989](#)) and one in rats with co-exposure with sulfur dioxide ([Laskin et al., 1970](#); see [RIVM, 1989](#)). In both studies malignant lung tumours were observed.

In recent bioassay inhalation studies with naphthalene, Fischer 344/N rats developed neuroblastomas of the nasal olfactory epithelium after being exposed in inhalation chambers to 0, 10, 30, or 60 ppm (80, 52, 157, or 314 mg/m³) for 6 hours per day, on 5 days per week, for 105 weeks ([NTP, 2000](#)). The observed rates in males were 0/49, 0/49, 4/48, and 3/48, respectively, and in females 0/49, 2/49, 3/49, and 12/49, respectively. In addition, adenomas of the nasal respiratory epithelium were observed in 0/49, 06/49, 8/48, and 15/48 males and in 0/49, 0/49, 4/49, and 2/49 females, respectively. In the study with B6C3F₁ mice subjected to whole-body exposure of 0, 10, or 30 ppm (0, 52, or 157 mg/m³) naphthalene in inhalation chambers for 6 hours per day, 5 days per week, for 104 weeks, a statistically significant increase in the incidence of bronchioloalveolar adenomas in high-dose female mice was observed ([NTP, 2000](#)). Increased incidences of bronchioloalveolar adenomas and carcinomas were observed in the male mice, but the increases were not statistically significant.

PAHs and their metabolites will also cause lung cancer in animals when administered by other routes. Classically, the newborn mouse model of lung cancer was used to rank the tumorigenicity of different B[a]P metabolites, given that the developing lung is more susceptible to carcinogen exposure. Studies such as these showed that the (+)-*anti*-B[a]PDE was the most potent lung tumorigen of the known B[a]P metabolites ([Buening et al., 1978](#);

[Kapitulnik et al., 1978](#)). Similarly, in the A/J mouse lung model of B[a]P-induced carcinogenesis, *anti*-B[a]PDE-DNA adducts were early lesions that could be detected in the initiation phase ([Nesnow et al., 1998](#)).

Carcinogenesis experiments with mixtures containing PAHs have also been reported. [Heinrich et al. \(1994\)](#) exposed groups of 72 female Wistar rats to a coal tar/pitch aerosol containing either 20 or 46 µg/m³ B[a]P for 17 hours per day, 5 days per week, for 10 or 20 months, followed by a clear air period of up to 20 or 10 months, respectively. The cumulative doses of inhaled B[a]P of the four exposure groups were 71, 143, 158, and 321 mg B[a]P/m³ hours, and the corresponding lung tumour rates were 4.2%, 33.3%, 38.9%, and 97.2%, respectively, whereas there were no tumours in the control group. In similar experiments in which rats were exposed to coal tar/pitch vapour condensed on the surface of fine carbon black particles, the resulting lung tumour rate was about twice as high.

[Pott and Heinrich \(1990\)](#) have also performed a lifelong inhalation study with rats exposed to diesel exhaust. In this study, tumour rates similar to those in the study with pitch pyrolysis vapours were induced, although the PAH content (measured as B[a]P) was 100–1000 times lower. This result indicates that diesel exhaust contains other potent carcinogenic or tumour-promoting compounds besides unsubstituted PAHs.

Numerous carcinogenicity studies have been performed using dermal application and subcutaneous and intramuscular injection (for overview, see [WHO, 1998](#)). An oral gavage study with B[a]P revealed tumour development in the liver, forestomach, auditory canal, oral cavity, skin, and intestines in both sexes of rats, and additionally the kidney in males and the mammary gland and oesophagus in females ([RIVM, 2001](#)). However, no lung tumours were observed after this route of administration. In a feeding study of B[a]P in mice, tumours in the tongue,

Table 7.5 Carcinogenicity of individual PAHs in experimental animals after inhalation or intratracheal instillation

Compound	Carcinogenicity (weight of evidence)	Species	No. of studies with positive, negative, and questionable results		
			+	-	±
Anthanthrene	Positive	Mouse	1		
Anthracene	Negative	Rat		1	
Benzo[<i>b</i>]fluoranthene	Positive	Rat Hamster	1	1	
Benzo[<i>j</i>]fluoranthene	Positive	Rat	1		
Benzo[<i>k</i>]fluoranthene	Positive	Rat	1		
Benzo[<i>g,h,i</i>]perylene	Negative	Rat		1	
Benzo[<i>a</i>]pyrene	Positive	Mouse Rat Hamster	1 9 11	1	1
Benzo[<i>e</i>]pyrene	Negative	Rat		1	
Chrysene	Positive	Rat	1		
Dibenz[<i>a,h</i>]anthracene	Positive	Rat Hamster	1 1	1	
Dibenzo[<i>a,i</i>]pyrene	Positive	Hamster	2		
Indeno[1,2,3- <i>cd</i>]pyrene	Positive	Rat	1		
Naphthalene	Positive	Mouse Rat	1		2
Phenanthrene	Negative	Rat		1	
Pyrene	Negative	Hamster		1	

Source: [WHO \(1998\)](#); reproduced with permission from the publisher; [IARC \(2002\)](#).

oesophagus, forestomach, and larynx, but not lung, were observed ([Culp et al., 1998](#)).

Carcinogenicity studies in humans

Occupational exposures

A review and meta-analysis on the association between occupational exposure to PAHs and lung cancer development in 39 cohorts found an average relative risk of 1.20 per 100 $\mu\text{g}/\text{m}^3$ years cumulative B[*a*]P ([Armstrong et al., 2004](#)). For some occupations relative risks were considerably higher, but confidence intervals were very wide. For exposures in coke ovens, gas works, and aluminium industries, the risk is equivalent to a relative risk of 1.06 for a working lifetime of 40 years at 1 $\mu\text{g}/\text{m}^3$.

Ambient air exposures

Few studies have addressed the impact of exposure to PAHs in ambient air on human cancer. Studies using other exposure indicators (PM or NO_2) have shown associations between air pollution and lung cancer; however, no PAH exposure information was available ([Pope et al., 2002](#); [Hoek et al., 2002](#); [Nafstad et al., 2003](#)). An analysis of the United States data on lung cancer, PM exposure, and older PAH and metal air concentration data, supports the plausibility that known chemical carcinogens may be responsible for the lung cancer attributed to $\text{PM}_{2.5}$ exposure in the American Cancer Society study ([Harrison et al., 2004](#)). A study by [Cordier et al. \(2004\)](#) found an increased risk of childhood brain cancer associated with PAH exposure. Both paternal

preconception occupational PAH exposure and paternal smoking were associated with increased risks for childhood brain tumours.

Human susceptibility

PAHs are metabolically activated by phase I P450 isozymes (*CYP1A1*, *CYP1B1*) in combination with epoxide hydrolase (*EPHX*) and phase I AKR isozymes (*AKR1A1*, *AKR1C1*-*AKR1C4*) and are detoxified by phase II enzymes including *GSTs*, *UTGs*, *SULTs*, and *COMT*. In addition, bulky covalent diol-epoxide DNA adducts can be repaired by nucleotide excision repair proteins (*XPD* [helicase], *XPA*, and *XPC* [damage recognition]), and oxidative DNA lesions can be repaired by base excision repair enzymes (*hOGG1* and *APE*). Each of these genes is highly polymorphic in the human population. (A complete list of these variants is available at the NCBI database: <http://www.ncbi.nlm.nih.gov/>.) Many of these variants are non-synonymous single-nucleotide polymorphisms (nSNPs) that can affect enzyme activity. Combinations of these nSNPs rather than an individual SNP may affect human genetic susceptibility to PAH emissions in ambient air.

In a study of Prague policemen occupationally exposed to polluted air, B[a]P-like DNA adducts were detected and found to be positively associated with SNPs in *XPD* and *GSTM1* ([Binková et al., 2007](#)). In another lung cancer case-control study, exposure to environmental tobacco smoke and polymorphisms in *CYP1B1* *Leu(432)Val* was significantly associated with lung cancer susceptibility, with an odds ratio for at least one allele of 2.87 (95% confidence interval [CI], 1.63–5.07) ([Wenzlaff et al., 2005a](#)). Combinations of the polymorphism in this phase I enzyme gene along with those selected from either phase II enzyme genes (*GSTM1* null, *GSTP1* *Ile(105)Val*) or NADPH-quinone oxidoreductase (*NQO1* *C(609)T*) were also evaluated. Here the combination of the *CYP1B1* *Leu(432)Val* allele and the *NQO1* *C(609)T* allele was associated with

the highest risk of lung cancer (odds ratio [OR], 4.14; 95% CI, 1.60–10.74) ([Wenzlaff et al., 2005a](#)). In the same study cohort, variants in *GSTM1*, *GSTT1*, and *GSTP1* were examined to determine whether there was an association of the genotype with lung cancer incidence in never-smokers. Individuals who had been exposed to household environmental tobacco smoke for > 20 years, and who were carriers of either the *GSTM1* null allele or the *GSTP1* Val allele, were at a 4-fold increased risk of developing lung cancer (OR, 4.56; 95% CI, 1.21–17.21) ([Wenzlaff et al., 2005b](#)). In a lung cancer case-control study in China, women who were never-smokers were found to be at a significant increased risk of adenocarcinoma if they were carriers of the variants in the nucleotide excision repair variant *XRCC1* 399 Gln/Gln versus the Arg/Arg genotype (OR, 14.12; 95% CI, 2.14–92.95). The OR of lung adenocarcinoma for the *XRCC1* 399Gln allele with exposure to cooking oil smoke was 6.29 (95% CI, 1.99–19.85) ([Li et al., 2005](#)). DNA integrity was investigated in 50 bus drivers, 20 garage men, and 50 controls in the Czech Republic and associated with variants in the base excision repair gene *hOGG1*. Carriers of at least one variant (Cys allele) had a higher degree of DNA damage ([Bagryantseva et al., 2010](#)). To date, no molecular epidemiological study has been performed whereby combinations of polymorphic variants in phase I, phase II, and DNA repair genes have been pooled. However, based on the studies described, carriers of variants in all three classes of genes might be at higher risk of developing lung cancer from emissions of PAHs in ambient air.

Conclusions

PAHs generated from the incomplete combustion of organic material are ubiquitous contaminants in urban air. There are numerous unsubstituted PAHs (pyrogenic) and substituted PAHs (petrogenic). The pyrogenic PAHs may occur in the gas phase, particulate phase,

or mixtures of both phases. The major world-wide source is the combustion of biofuels, while other sources such as combustion plants, various industrial and production processes, road transport, and waste incineration can contribute. Total PAH levels in some urban areas are in the range of 100–200 ng/m³ but may be even higher in more polluted areas and can show distinct seasonal variation. However, measurements of total PAHs are relatively scarce. B[a]P is the traditional marker for PAHs, but various other individual PAHs have also been proposed, such as fluoranthene, B[a]P, and benzo[b]fluoranthene. Biomarkers of exposure include 1-hydroxypyrene, 3-hydroxy-B[a]P, and tetraols, but DNA and protein adducts can also be measured as intermediate cancer biomarkers. The major disease end-point of interest is lung cancer, and approximately 10–15% of all lung cancer cases are seen in never-smokers. Parent PAHs must be metabolically activated to electrophilic intermediates (radical cations, vicinal diol-epoxides, and *o*-quinones) to act as lung carcinogens. All three routes have been observed in human lung cells. Various promotional effects of PAHs may contribute to their carcinogenic action. In all, 10 PAHs have been found to be carcinogenic in experimental animals after inhalation or intratracheal instillation. Naphthalene seems to be an exception compared with other carcinogenic PAHs as it appears to not be genotoxic. A meta-analysis of occupational cohort studies found a 20% increase in relative risk per 100 µg/m³ years cumulative B[a]P exposure. Studies of ambient air pollution and cancer have demonstrated an association between carriers of polymorphic variants in phase I, phase II, and DNA repair enzyme genes.

References

- Abbas I, Garcon G, Saint-Georges F *et al.* (2013). Polycyclic aromatic hydrocarbons within airborne particulate matter (PM 2.5) produced DNA bulky stable adducts in a human lung cell coculture model. *J Appl Toxicol*, 33: 109–119. PMID:[21913209](#)
- Armstrong B, Hutchinson E, Unwin J, Fletcher T (2004). Lung cancer risk after exposure to polycyclic aromatic hydrocarbons: a review and meta-analysis. *Environ Health Perspect*, 112: 970–978. doi:[10.1289/ehp.6895](#) PMID:[15198916](#)
- Bagryantseva Y, Novotna B, Rossner P Jr *et al.* (2010). Oxidative damage to biological macromolecules in Prague bus drivers and garagemen: impact of air pollution and genetic polymorphisms. *Toxicol Lett*, 199: 60–68. doi:[10.1016/j.toxlet.2010.08.007](#) PMID:[20723587](#)
- Benbrahim-Tallaa L, Baan RA, Grosse Y *et al.* (2012). International Agency for Research on Cancer Monograph Working Group. Carcinogenicity of diesel-engine and gasoline-engine exhausts and some nitroarenes. *Lancet Oncol*, 13: 663–664. doi:[10.1016/S1470-2045\(12\)70280-2](#) PMID:[22946126](#)
- Binková B, Cerná M, Pastorková A *et al.* (2003). Biological activities of organic compounds adsorbed onto ambient air particles: comparison between the cities of Teplice and Prague during the summer and winter seasons 2000–2001. *Mutat Res*, 525: 43–59. doi:[10.1016/S0027-5107\(02\)00312-3](#) PMID:[12650904](#)
- Binková B, Chvatalova I, Lnenickova Z *et al.* (2007). PAH-DNA adducts in environmentally exposed population in relation to metabolic and DNA repair gene polymorphisms. *Mutat Res*, 620: 49–61. doi:[10.1016/j.mrfmmm.2007.02.022](#) PMID:[17412371](#)
- Bond JA, Harkema JR, Russell VI (1988). Regional distribution of xenobiotic metabolizing enzymes in respiratory airways of dogs. *Drug Metab Dispos*, 16: 116–124. PMID:[2894939](#)
- Boström CE, Gerde P, Hanberg A *et al.* (2002). Cancer risk assessment, indicators, and guidelines for polycyclic aromatic hydrocarbons in the ambient air. *Environ Health Perspect*, 110: Suppl 3: 451–488. doi:[10.1289/ehp.02110s3451](#) PMID:[12060843](#)
- Buening MK, Wislocki PG, Levin W *et al.* (1978). Tumorigenicity of the optical enantiomers of the diastereomeric benzo[a]pyrene 7,8-diol-9,10-epoxides in newborn mice: exceptional activity of (+)-7β,8α-dihydroxy-9α,10α-epoxy-7,8,9,10-tetrahydrobenzo[a]pyrene. *Proc Natl Acad Sci USA*, 75: 5358–5361. doi:[10.1073/pnas.75.11.5358](#) PMID:[281685](#)
- Burczynski ME, Lin HK, Penning TM (1999). Isoform-specific induction of a human aldo-keto reductase by polycyclic aromatic hydrocarbons (PAHs), electrophiles, and oxidative stress: implications for the

- alternative pathway of PAH activation catalyzed by human dihydrodiol dehydrogenase. *Cancer Res*, 59: 607–614. PMID:[9973208](#)
- Burczynski ME & Penning TM (2000). Genotoxic polycyclic aromatic hydrocarbon *ortho*-quinones generated by aldo-keto reductases induce CYP1A1 via nuclear translocation of the aryl hydrocarbon receptor. *Cancer Res*, 60: 908–915. PMID:[10706104](#)
- Burdick AD, Davis JW 2nd, Liu KJ *et al.* (2003). Benzo(a) pyrene quinones increase cell proliferation, generate reactive oxygen species, and transactivate the epidermal growth factor receptor in breast epithelial cells. *Cancer Res*, 63: 7825–7833. PMID:[14633709](#)
- Casale GP, Higginbotham S, Johansson SL *et al.* (1997). Inflammatory response of mouse skin exposed to the very potent carcinogen dibenzo[*a,l*]pyrene: a model for tumor promotion. *Fundam Appl Toxicol*, 36: 71–78. doi:[10.1006/faat.1997.2291](#) PMID:[9073469](#)
- Casale GP, Singhal M, Bhattacharya S *et al.* (2001). Detection and quantification of depurinated benzo[*a*] pyrene-adducted DNA bases in the urine of cigarette smokers and women exposed to household coal smoke. *Chem Res Toxicol*, 14: 192–201. doi:[10.1021/tx000012y](#) PMID:[11258968](#)
- Cavaliere EL, Higginbotham S RamaKrishna NVS *et al.* (1991). Comparative dose-response tumorigenicity studies of dibenzo[*alpha,l*]pyrene versus 7,12-dimethylbenz[*alpha*]anthracene, benzo[*alpha*]pyrene and two dibenzo[*alpha,l*]pyrene dihydrodiols in mouse skin and rat mammary gland. *Carcinogenesis*, 12:1939–1944. doi:[10.1093/carcin/12.10.1939](#) PMID:[1934274](#)
- Cavaliere EL & Rogan EG (1995). Central role of radical cations in metabolic activation of polycyclic aromatic hydrocarbons. *Xenobiotica*, 25: 677–688. doi:[10.3109/00498259509061885](#) PMID:[7483666](#)
- Cavaliere EL, Rogan EG, Devanesan PD *et al.* (1990). Binding of benzo[*a*]pyrene to DNA by cytochrome P-450 catalyzed one-electron oxidation in rat liver microsomes and nuclei. *Biochemistry*, 29: 4820–4827. doi:[10.1021/bi00472a011](#) PMID:[2364062](#)
- Cavaliere EL, Rogan EG, Li K-M *et al.* (2005). Identification and quantification of the depurinating DNA adducts formed in mouse skin treated with dibenzo[*a,l*]pyrene (DB[*a,l*]P) or its metabolites and in rat mammary gland treated with DB[*a,l*]P. *Chem Res Toxicol*, 18: 976–983. doi:[10.1021/tx049682k](#) PMID:[15962932](#)
- Chen RH, Maher VM, McCormick JJ (1990). Effect of excision repair by diploid human fibroblasts on the kinds and locations of mutations induced by (+/-)-7 β ,8 α -dihydroxy-9 α ,10 α -epoxy-7,8,9,10-tetrahydrobenzo[*a*]pyrene in the coding region of the HPRT gene. *Proc Natl Acad Sci USA*, 87: 8680–8684. doi:[10.1073/pnas.87.21.8680](#) PMID:[2122466](#)
- Conney AH (1982). Induction of microsomal enzymes by foreign chemicals and carcinogenesis by polycyclic aromatic hydrocarbons: G. H. A. Clowes Memorial Lecture. *Cancer Res*, 42: 4875–4917. PMID:[6814745](#)
- Constantin D, Mehrotra K, Rahimtula A *et al.* (1994). Stimulatory effects of sulfur and nitrogen oxides on carcinogen activation in human polymorphonuclear leukocytes. *Environ Health Perspect*, 102: Suppl 4: 161–164. PMID:[7821291](#)
- Cooper CS, Ribeiro O, Farmer PB *et al.* (1980). The metabolic activation of benz[*a*]anthracene in hamster embryo cells: evidence that diol-epoxides react with guanosine, deoxyguanosine and adenosine in nucleic acids. *Chem Biol Interact*, 32: 209–231. doi:[10.1016/0009-2797\(80\)90079-4](#) PMID:[6159115](#)
- Cordier S, Monfort C, Filippini G *et al.* (2004). Parental exposure to polycyclic aromatic hydrocarbons and the risk of childhood brain tumors: The SEARCH International Childhood Brain Tumor Study. *Am J Epidemiol*, 159: 1109–1116. doi:[10.1093/aje/kwh154](#) PMID:[15191928](#)
- CORINAIR (1997). *EMEP/CORINAIR Emission Inventory Guidebook - 2007*. Technical Report No. 16/2007. Copenhagen: European Environmental Agency.
- Culp SJ, Gaylor DW, Sheldon WG *et al.* (1998). A comparison of the tumors induced by coal tar and benzo[*a*]pyrene in a 2-year bioassay. *Carcinogenesis*, 19: 117–124. doi:[10.1093/carcin/19.1.117](#) PMID:[9472702](#)
- Day BW, Naylor S, Gan LS *et al.* (1990). Molecular dosimetry of polycyclic aromatic hydrocarbon epoxides and diol epoxides via hemoglobin adducts. *Cancer Res*, 50: 4611–4618. PMID:[2369737](#)
- Denison MS, Fisher JM, Whitlock JP Jr (1988b). Inducible, receptor-dependent protein-DNA interactions at a dioxin-responsive transcriptional enhancer. *Proc Natl Acad Sci USA*, 85: 2528–2532. doi:[10.1073/pnas.85.8.2528](#) PMID:[2833743](#)
- Denison MS, Fisher JM, Whitlock JP Jr (1989). Protein-DNA interactions at recognition sites for the dioxin-Ah receptor complex. *J Biol Chem*, 264: 16478–16482. PMID:[2550446](#)
- Denison MS, Fisher JM, Whitlock JP Jr (1988a). The DNA recognition site for the dioxin-Ah receptor complex. Nucleotide sequence and functional analysis. *J Biol Chem*, 263: 17221–17224. PMID:[2846558](#)
- Denissenko MF, Pao A, Tang M, Pfeifer GP (1996). Preferential formation of benzo[*a*]pyrene adducts at lung cancer mutational hotspots in P53. *Science*, 274: 430–432. doi:[10.1126/science.274.5286.430](#) PMID:[8832894](#)
- Fan R, Wang D, Mao C *et al.* (2012). Preliminary study of children's exposure to PAHs and its association with 8-hydroxy-2'-deoxyguanosine in Guangzhou, China. *Environ Int*, 42: 53–58. doi:[10.1016/j.envint.2011.03.021](#) PMID:[21511339](#)
- Gaga EO, Ari A, Döğeroğlu T *et al.* (2012). Atmospheric polycyclic aromatic hydrocarbons in an industrialized city, Kocaeli, Turkey: study of seasonal variations,

- influence of meteorological parameters and health risk estimation. *J Environ Monit*, 14: 2219–2229. doi:[10.1039/c2em30118k](https://doi.org/10.1039/c2em30118k) PMID:[22699796](https://pubmed.ncbi.nlm.nih.gov/22699796/)
- Geacintov NE, Cosman M, Hingerty BE *et al.* (1997). NMR solution structures of stereoisometric covalent polycyclic aromatic carcinogen-DNA adduct: principles, patterns, and diversity. *Chem Res Toxicol*, 10: 111–146. doi:[10.1021/tx9601418](https://doi.org/10.1021/tx9601418) PMID:[9049424](https://pubmed.ncbi.nlm.nih.gov/9049424/)
- Gerde P, Muggenburg BA, Hoover MD, Henderson RF (1993). Disposition of polycyclic aromatic hydrocarbons in the respiratory tract of the beagle dog. I. The alveolar region. *Toxicol Appl Pharmacol*, 121: 313–318. doi:[10.1006/taap.1993.1159](https://doi.org/10.1006/taap.1993.1159) PMID:[8346548](https://pubmed.ncbi.nlm.nih.gov/8346548/)
- Glatt H, Piée A, Pauly K *et al.* (1991). Fjord- and bay-region diol-epoxides investigated for stability, SOS induction in *Escherichia coli*, and mutagenicity in *Salmonella typhimurium* and mammalian cells. *Cancer Res*, 51: 1659–1667. PMID:[1900215](https://pubmed.ncbi.nlm.nih.gov/1900215/)
- Gräslund A & Jernström B (1989). DNA-carcinogen interaction: covalent DNA-adducts of benzo(a)pyrene 7,8-dihydrodiol 9,10-epoxides studied by biochemical and biophysical techniques. *Q Rev Biophys*, 22: 1–37. PMID:[2501820](https://pubmed.ncbi.nlm.nih.gov/2501820/)
- Guengerich FP (1993). The 1992 Bernard B. Brodie Award Lecture. Bioactivation and detoxication of toxic and carcinogenic chemicals. *Drug Metab Dispos*, 21: 1–6. PMID:[8095200](https://pubmed.ncbi.nlm.nih.gov/8095200/)
- Hainaut P & Pfeifer GP (2001). Patterns of p53 G→T transversions in lung cancers reflect the primary mutagenic signature of DNA-damage by tobacco smoke. *Carcinogenesis*, 22: 367–374. doi:[10.1093/carcin/22.3.367](https://doi.org/10.1093/carcin/22.3.367) PMID:[11238174](https://pubmed.ncbi.nlm.nih.gov/11238174/)
- Hansen LD, Nestler C, Ringelberg D, Bajpai R (2004). Extended bioremediation of PAH/PCP contaminated soils from the POPILE wood treatment facility. *Chemosphere*, 54: 1481–1493. doi:[10.1016/j.chemosphere.2003.09.046](https://doi.org/10.1016/j.chemosphere.2003.09.046) PMID:[14659950](https://pubmed.ncbi.nlm.nih.gov/14659950/)
- Harrison RM, Smith DJT, Kibble AJ (2004). What is responsible for the carcinogenicity of PM_{2.5}? *Occup Environ Med*, 61: 799–805. doi:[10.1136/oem.2003.010504](https://doi.org/10.1136/oem.2003.010504) PMID:[15377764](https://pubmed.ncbi.nlm.nih.gov/15377764/)
- Hecht SS, Carmella SG, Villalta PW, Hochalter JB (2010). Analysis of phenanthrene and benzo(a)pyrene tetraol enantiomers in human urine: relevance to the bay region diol epoxide hypothesis of benzo(a)pyrene carcinogenesis and to biomarker studies. *Chem Res Toxicol*, 23: 900–908. doi:[10.1021/tx9004538](https://doi.org/10.1021/tx9004538) PMID:[20369855](https://pubmed.ncbi.nlm.nih.gov/20369855/)
- Heinrich U, Roller M, Pott F (1994). Estimation of a lifetime unit lung cancer risk for benzo(a)pyrene based on tumour rates in rats exposed to coal tar/pitch condensation aerosol. *Toxicol Lett*, 72: 155–161. doi:[10.1016/0378-4274\(94\)90023-X](https://doi.org/10.1016/0378-4274(94)90023-X) PMID:[8202928](https://pubmed.ncbi.nlm.nih.gov/8202928/)
- Hoek G, Brunekreef B, Goldbohm S *et al.* (2002). Association between mortality and indicators of traffic-related air pollution in the Netherlands: a cohort study. *Lancet*, 360: 1203–1209. doi:[10.1016/S0140-6736\(02\)11280-3](https://doi.org/10.1016/S0140-6736(02)11280-3) PMID:[12401246](https://pubmed.ncbi.nlm.nih.gov/12401246/)
- Hoh E, Hunt RN, Quintana PJ *et al.* (2012). Environmental tobacco smoke as a source of polycyclic aromatic hydrocarbons in settled household dust. *Environ Sci Technol*, 46: 4174–4183. doi:[10.1021/es300267g](https://doi.org/10.1021/es300267g) PMID:[22397504](https://pubmed.ncbi.nlm.nih.gov/22397504/)
- IARC (1989). Occupational exposures in petroleum refining; crude oil and major petroleum fuels. *IARC Monogr Eval Carcinog Risks Hum*, 45: 1–322. PMID:[2664246](https://pubmed.ncbi.nlm.nih.gov/2664246/)
- IARC (1983). Polynuclear aromatic compounds, Part 1, chemical, environmental and experimental data. *IARC Monogr Eval Carcinog Risk Chem Hum*, 32: 1–453. PMID:[6586639](https://pubmed.ncbi.nlm.nih.gov/6586639/)
- IARC (1984). Polynuclear aromatic compounds, Part 3, industrial exposures in aluminium production, coal gasification, coke production, and iron and steel founding. *IARC Monogr Eval Carcinog Risk Chem Hum*, 34: 1–219.
- IARC (1985). Polynuclear aromatic compounds, Part 4, bitumens, coal-tars and derived products, shale-oils and soots. *IARC Monogr Eval Carcinog Risk Chem Hum*, 35: 1–247. PMID:[2991123](https://pubmed.ncbi.nlm.nih.gov/2991123/)
- IARC (1984). Polynuclear aromatic hydrocarbons, Part 2, carbon blacks, mineral oils (lubricant base oils and derived products) and some nitroarenes. *IARC Monogr Eval Carcinog Risk Chem Hum*, 33: 1–222. PMID:[6590450](https://pubmed.ncbi.nlm.nih.gov/6590450/)
- IARC (2010). Some non-heterocyclic polycyclic aromatic hydrocarbons and some related exposures. *IARC Monogr Eval Carcinog Risks Hum*, 92: 1–853. PMID:[21141735](https://pubmed.ncbi.nlm.nih.gov/21141735/)
- IARC (2002). Some traditional herbal medicines, some mycotoxins, naphthalene and styrene. *IARC Monogr Eval Carcinog Risks Hum*, 82: 1–556. PMID:[12687954](https://pubmed.ncbi.nlm.nih.gov/12687954/)
- Jeffrey AM (1985). Polycyclic aromatic hydrocarbon-DNA adducts. Formation, detection, and characterization. In: Harvey RG, ed. *Polycyclic Hydrocarbons and Carcinogenesis*. Washington, DC: American Chemical Society, pp. 187–208.
- Jerina DM, Chadha A, Cheh AM *et al.* (1991). Covalent bonding of bay-region diol epoxides to nucleic acids. *Adv Exp Med Biol*, 283: 533–553. doi:[10.1007/978-1-4684-5877-0_70](https://doi.org/10.1007/978-1-4684-5877-0_70) PMID:[2069024](https://pubmed.ncbi.nlm.nih.gov/2069024/)
- Jin Y & Penning TM (2007). Aldo-keto reductases and bioactivation/detoxication. *Annu Rev Pharmacol Toxicol*, 47: 263–292. doi:[10.1146/annurev.pharmtox.47.120505.105337](https://doi.org/10.1146/annurev.pharmtox.47.120505.105337) PMID:[16970545](https://pubmed.ncbi.nlm.nih.gov/16970545/)
- Jung KH, Yan B, Chillrud SN *et al.* (2010). Assessment of benzo(a)pyrene-equivalent carcinogenicity and mutagenicity of residential indoor versus outdoor polycyclic aromatic hydrocarbons exposing young children in New York City. *Int J Environ Res Public Health*, 7: 1889–1900. doi:[10.3390/ijerph7051889](https://doi.org/10.3390/ijerph7051889) PMID:[20622999](https://pubmed.ncbi.nlm.nih.gov/20622999/)
- Kapitulnik J, Wislocki PG, Levin W *et al.* (1978). Tumorigenicity studies with diol-epoxides of benzo(a)

- pyrene which indicate that (+/-)-*trans*-7 β ,8 α -dihydroxy-9 α ,10 α -epoxy-7,8,9,10-tetrahydrobenzo(a)pyrene is an ultimate carcinogen in newborn mice. *Cancer Res*, 38: 354–358. PMID:[620406](#)
- Koehl W, Amin S, Staretz ME *et al.* (1996). Metabolism of 5-methylchrysene and 6-methylchrysene by human hepatic and pulmonary cytochrome P450 enzymes. *Cancer Res*, 56: 316–324. PMID:[8542586](#)
- Li MC, Cui ZS, He QC, Zhou BS (2005). Association of genetic polymorphism in the DNA repair gene XRCC1 with susceptibility to lung cancer in non-smoking women [in Chinese]. *Zhonghua Zhong Liu Za Zhi*, 27: 713–716. PMID:[16483478](#)
- Li Z, Sjödin A, Romanoff LC *et al.* (2011). Evaluation of exposure reduction to indoor air pollution in stove intervention projects in Peru by urinary biomonitoring of polycyclic aromatic hydrocarbon metabolites. *Environ Int*, 37: 1157–1163. doi:[10.1016/j.envint.2011.03.024](#) PMID:[21524795](#)
- Lu D, Harvey RG, Blair IA, Penning TM (2011). Quantitation of benzo[a]pyrene metabolic profiles in human bronchoalveolar (H358) cells by stable isotope dilution liquid chromatography-atmospheric pressure chemical ionization mass spectrometry. *Chem Res Toxicol*, 24: 1905–1914. doi:[10.1021/tx2002614](#) PMID:[21962213](#)
- Luch A, Schober W, Soballa VJ *et al.* (1999). Metabolic activation of dibenzo[a,l]pyrene by human cytochrome P450 1A1 and P450 1B1 expressed in V79 Chinese hamster. *Chem Res Toxicol*, 12: 353–364. doi:[10.1021/tx980240g](#) PMID:[10207125](#)
- Luch A, Seidel A, Glatt H, Platt KL (1997). Metabolic activation of the (+)-S,S- and (-)-R,R-enantiomers of *trans*-11,12-dihydroxy-11,12-dihydrodibenzo[a,l]pyrene: stereoselectivity, DNA adduct formation, and mutagenicity in Chinese hamster V79 cells. *Chem Res Toxicol*, 10: 1161–1170. doi:[10.1021/tx970005i](#) PMID:[9348439](#)
- MacLeod MC, Adair G, Humphrey RM (1988). Differential efficiency of mutagenesis at three genetic loci in CHO cells by a benzo[a]pyrene diol epoxide. *Mutat Res*, 199: 243–254. PMID:[3129654](#)
- Malaveille C, Kuroki T, Sims P *et al.* (1977). Mutagenicity of isomeric diol-epoxides of benzo[a]pyrene and benz[a]anthracene in *S. typhimurium* TA98 and TA100 and in V79 Chinese hamster cells. *Mutat Res*, 44: 313–326. doi:[10.1016/0027-5107\(77\)90091-4](#) PMID:[333280](#)
- Marshall CJ, Vousden KH, Phillips DH (1984). Activation of c-Ha-ras-1 proto-oncogene by *in vitro* modification with a chemical carcinogen, benzo(a)pyrene diol epoxide. *Nature*, 310: 586–589. doi:[10.1038/310586a0](#) PMID:[6431299](#)
- Meijer SN, Sweetman AJ, Halsall CJ, Jones KC (2008). Temporal trends of polycyclic aromatic hydrocarbons in the U.K. atmosphere: 1991–2005. *Environ Sci Technol*, 42: 3213–3218. doi:[10.1021/es702979d](#) PMID:[18522096](#)
- Melikian AA, LaVoie EJ, Hecht SS, Hoffmann D (1983). 5-Methylchrysene metabolism in mouse epidermis *in vivo*, diol epoxide–DNA adduct persistence, and diol epoxide reactivity with DNA as potential factors influencing the predominance of 5-methylchrysene-1,2-diol-3,4-epoxide–DNA adducts in mouse epidermis. *Carcinogenesis*, 4: 843–849. doi:[10.1093/carcin/4.7.843](#) PMID:[6872139](#)
- Nafstad P, Håheim LL, Oftedal B *et al.* (2003). Lung cancer and air pollution: a 27 year follow up of 16 209 Norwegian men. *Thorax*, 58: 1071–1076. doi:[10.1136/thorax.58.12.1071](#) PMID:[14645978](#)
- Nebert DW, Dalton TP, Okey AB, Gonzalez FJ (2004). Role of aryl hydrocarbon receptor-mediated induction of the CYP1 enzymes in environmental toxicity and cancer. *J Biol Chem*, 279: 23847–23850. doi:[10.1074/jbc.R400004200](#) PMID:[15028720](#)
- Nebert DW & Jensen NM (1979). The Ah locus: genetic regulation of the metabolism of carcinogens, drugs, and other environmental chemicals by cytochrome P-450-mediated monooxygenases. *CRC Crit Rev Biochem*, 6: 401–437. doi:[10.3109/10409237909105427](#) PMID:[378536](#)
- Nebert DW, Puga A, Vasiliou V (1993). Role of the Ah receptor and the dioxin-inducible [Ah] gene battery in toxicity, cancer, and signal transduction. *Ann N Y Acad Sci*, 685: 624–640. doi:[10.1111/j.1749-6632.1993.tb35928.x](#) PMID:[8395783](#)
- Nesnow S, Mass MJ, Ross JA *et al.* (1998). Lung tumorigenic interactions in strain A/J mice of five environmental polycyclic aromatic hydrocarbons. *Environ Health Perspect*, 106: Suppl 6: 1337–1346. doi:[10.1289/ehp.98106s61337](#) PMID:[9860890](#)
- NTP (2000). *Toxicology and Carcinogenesis Studies of Naphthalene (CAS No. 91–20–3) in F344/N Rats (Inhalation Studies)*. National Toxicology Program Technical Report No. 500; NIH Publ. No. 01-4434, Research Triangle Park, NC.
- Palackal NT, Burczynski ME, Harvey RG, Penning TM (2001). The ubiquitous aldehyde reductase (AKR1A1) oxidizes proximate carcinogen *trans*-dihydrodiols to *o*-quinones: potential role in polycyclic aromatic hydrocarbon activation. *Biochemistry*, 40: 10901–10910. doi:[10.1021/bi010872t](#) PMID:[11535067](#)
- Palackal NT, Lee S-H, Harvey RG *et al.* (2002). Activation of polycyclic aromatic hydrocarbon *trans*-dihydrodiol proximate carcinogens by human aldo-keto reductase (AKR1C) enzymes and their functional overexpression in human lung carcinoma (A549) cells. *J Biol Chem*, 277: 24799–24808. doi:[10.1074/jbc.M112424200](#) PMID:[11978787](#)
- Park J-H, Gelhaus S, Vedantam S *et al.* (2008a). The pattern of p53 mutations caused by PAH *o*-quinones is driven by 8-oxo-dGuo formation while the spectrum of mutations is determined by biological selection

- for dominance. *Chem Res Toxicol*, 21: 1039–1049. doi:[10.1021/tx700404a](https://doi.org/10.1021/tx700404a) PMID:[18489080](https://pubmed.ncbi.nlm.nih.gov/18489080/)
- Park J-H, Mangal D, Tacka KA *et al.* (2008b). Evidence for the aldo-keto reductase pathway of polycyclic aromatic *trans*-dihydrodiol activation in human lung A549 cells. *Proc Natl Acad Sci USA*, 105: 6846–6851. doi:[10.1073/pnas.0802776105](https://doi.org/10.1073/pnas.0802776105) PMID:[18474869](https://pubmed.ncbi.nlm.nih.gov/18474869/)
- Park J-H, Penning TM (2008). Polyaromatic hydrocarbons. In: Stadler RH, Lineback DR, eds. *Process-Induced Food Toxicants: Occurrence, Formation, Mitigation, and Health Risks*, Chapter 2. Hoboken, NJ: John Wiley & Sons, Inc. doi:[10.1002/9780470430101.ch2h](https://doi.org/10.1002/9780470430101.ch2h)
- Penning TM (2004). Aldo-keto reductases and formation of polycyclic aromatic hydrocarbon *o*-quinones. *Methods Enzymol*, 378: 31–67. doi:[10.1016/S0076-6879\(04\)78003-9](https://doi.org/10.1016/S0076-6879(04)78003-9) PMID:[15038957](https://pubmed.ncbi.nlm.nih.gov/15038957/)
- Penning TM, Burczynski ME, Hung C-F *et al.* (1999). Dihydrodiol dehydrogenases and polycyclic aromatic hydrocarbon activation: generation of reactive and redox active *o*-quinones. *Chem Res Toxicol*, 12: 1–18. doi:[10.1021/tx980143n](https://doi.org/10.1021/tx980143n) PMID:[9894013](https://pubmed.ncbi.nlm.nih.gov/9894013/)
- Penning TM & Drury JE (2007). Human aldo-keto reductases: Function, gene regulation, and single nucleotide polymorphisms. *Arch Biochem Biophys*, 464: 241–250. doi:[10.1016/j.abb.2007.04.024](https://doi.org/10.1016/j.abb.2007.04.024) PMID:[17537398](https://pubmed.ncbi.nlm.nih.gov/17537398/)
- Penning TM, Ohnishi ST, Ohnishi T, Harvey RG (1996). Generation of reactive oxygen species during the enzymatic oxidation of polycyclic aromatic hydrocarbon *trans*-dihydrodiols catalyzed by dihydrodiol dehydrogenase. *Chem Res Toxicol*, 9: 84–92. doi:[10.1021/tx950055s](https://doi.org/10.1021/tx950055s) PMID:[8924621](https://pubmed.ncbi.nlm.nih.gov/8924621/)
- Petruska JM, Mosebrook DR, Jakab GJ, Trush MA (1992). Myeloperoxidase-enhanced formation of (+)-*trans*-7,8-dihydroxy-7,8-dihydrobenzo[*a*]pyrene-DNA adducts in lung tissue *in vitro*: a role of pulmonary inflammation in the bioactivation of a procarcinogen. *Carcinogenesis*, 13: 1075–1081. doi:[10.1093/carcin/13.7.1075](https://doi.org/10.1093/carcin/13.7.1075) PMID:[1322250](https://pubmed.ncbi.nlm.nih.gov/1322250/)
- Platt KL, Schollmeier M, Frank H, Oesch F (1990). Stereoselective metabolism of dibenz(*a,h*)anthracene to *trans*-dihydrodiols and their activation to bacterial mutagens. *Environ Health Perspect*, 88: 37–41. doi:[10.1289/ehp.908837](https://doi.org/10.1289/ehp.908837) PMID:[2272331](https://pubmed.ncbi.nlm.nih.gov/2272331/)
- Pope CA 3rd, Burnett RT, Thun MJ *et al.* (2002). Lung cancer, cardiopulmonary mortality, and long-term exposure to fine particulate air pollution. *JAMA*, 287: 1132–1141. doi:[10.1001/jama.287.9.1132](https://doi.org/10.1001/jama.287.9.1132) PMID:[11879110](https://pubmed.ncbi.nlm.nih.gov/11879110/)
- Pott F, Heinrich U (1990). Relative significance of different hydrocarbons for the carcinogenic potency of emission from various incomplete combustion processes. In: Vainio H, Sorsa M, McMichael AJ, eds. *Complex Mixtures and Cancer Risk*. Lyon: International Agency for Research on Cancer, pp. 288–297.
- Prevedouros K, Brorström-Lundén E, Halsall CJ *et al.* (2004). Seasonal and long-term trends in atmospheric PAH concentrations: evidence and implications. *Environ Pollut*, 128: 17–27. PMID:[14667717](https://pubmed.ncbi.nlm.nih.gov/14667717/)
- RIVM (1989). *Integrated Criteria Document PAHs*. (RIVM Report No. 758474011). Bilthoven: National Institute of Public Health and the Environment.
- RIVM (2001). *Tumorigenic Effects in Wistar Rats Orally Administered Benzo[*a*]pyrene for Two Years (Gavage Studies). Implications for Human Cancer Risks Associated with Oral Exposure to Polycyclic Aromatic Hydrocarbons* (RIVM Report No. 658603 010). Bilthoven: National Institute of Public Health and the Environment.
- Ross JA, Nelson GB, Holden KL *et al.* (1992). DNA adducts and induction of sister chromatid exchanges in the rat following benzo[*b*]fluoranthene administration. *Carcinogenesis*, 13: 1731–1734. doi:[10.1093/carcin/13.10.1731](https://doi.org/10.1093/carcin/13.10.1731) PMID:[1423831](https://pubmed.ncbi.nlm.nih.gov/1423831/)
- Santella RM (1999). Immunological methods for detection of carcinogen-DNA damage in humans. *Cancer Epidemiol Biomarkers Prev*, 8: 733–739. PMID:[10498391](https://pubmed.ncbi.nlm.nih.gov/10498391/)
- Schauer C, Niessner R, Pöschl U (2004). Analysis of nitrated polycyclic aromatic hydrocarbons by liquid chromatography with fluorescence and mass spectrometry detection: air particulate matter, soot, and reaction product studies. *Anal Bioanal Chem*, 378: 725–736. doi:[10.1007/s00216-003-2449-1](https://doi.org/10.1007/s00216-003-2449-1) PMID:[14704835](https://pubmed.ncbi.nlm.nih.gov/14704835/)
- Shimada T, Gillam EMJ, Oda Y *et al.* (1999). Metabolism of benzo[*a*]pyrene to *trans*-7,8-dihydroxy-7,8-dihydrobenzo[*a*]pyrene by recombinant human cytochrome P450 1B1 and purified liver epoxide hydrolase. *Chem Res Toxicol*, 12: 623–629. doi:[10.1021/tx990028s](https://doi.org/10.1021/tx990028s) PMID:[10409402](https://pubmed.ncbi.nlm.nih.gov/10409402/)
- Shimada T, Oda Y, Gillam EMJ *et al.* (2001). Metabolic activation of polycyclic aromatic hydrocarbons and other procarcinogens by cytochromes P450 1A1 and P450 1B1 allelic variants and other human cytochromes P450 in *Salmonella typhimurium* NM2009. *Drug Metab Dispos*, 29: 1176–1182. PMID:[11502724](https://pubmed.ncbi.nlm.nih.gov/11502724/)
- Shultz CA, Quinn AM, Park JH *et al.* (2011). Specificity of human aldo-keto reductases, NAD(P)H:quinone oxidoreductase, and carbonyl reductases to redox-cycle polycyclic aromatic hydrocarbon diones and 4-hydroxyequilenin-*o*-quinone. *Chem Res Toxicol*, 24: 2153–2166. doi:[10.1021/tx200294c](https://doi.org/10.1021/tx200294c) PMID:[21910479](https://pubmed.ncbi.nlm.nih.gov/21910479/)
- Sims P & Grover PL (1974). Epoxides in polycyclic aromatic hydrocarbon metabolism and carcinogenesis. *Adv Cancer Res*, 20: 165–274. doi:[10.1016/S0065-230X\(08\)60111-6](https://doi.org/10.1016/S0065-230X(08)60111-6) PMID:[4617500](https://pubmed.ncbi.nlm.nih.gov/4617500/)
- Solhaug A, Refsnes M, Låg M *et al.* (2004). Polycyclic aromatic hydrocarbons induce both apoptotic and anti-apoptotic signals in Hepa1c1c7 cells. *Carcinogenesis*, 25: 809–819. doi:[10.1093/carcin/bgh069](https://doi.org/10.1093/carcin/bgh069) PMID:[14729587](https://pubmed.ncbi.nlm.nih.gov/14729587/)
- Suwan-ampai P, Navas-Acien A, Strickland PT, Agnew J (2009). Involuntary tobacco smoke exposure and urinary levels of polycyclic aromatic hydrocarbons

- in the United States, 1999 to 2002. *Cancer Epidemiol Biomarkers Prev*, 18: 884–893. doi:[10.1158/1055-9965.EPI-08-0939](https://doi.org/10.1158/1055-9965.EPI-08-0939) PMID:[19258471](https://pubmed.ncbi.nlm.nih.gov/19258471/)
- Thakker DR, Yagi H, Levin W *et al.* (1985). Polycyclic aromatic hydrocarbons: metabolic activation to ultimate carcinogens. In: Anders MW, ed. *Bioactivation of Foreign Compounds*. New York: Academic Press, pp. 177–2424.
- Thyssen J, Althoff J, Kimmerle G, Mohr U (1981). Inhalation studies with benzo[a]pyrene in Syrian golden hamsters. *J Natl Cancer Inst*, 66: 575–577. PMID:[6937711](https://pubmed.ncbi.nlm.nih.gov/6937711/)
- Topinka J, Sevastyanova O, Binková B *et al.* (2007). Biomarkers of air pollution exposure—a study of policemen in Prague. *Mutat Res*, 624: 9–17. doi:[10.1016/j.mrfmmm.2007.02.032](https://doi.org/10.1016/j.mrfmmm.2007.02.032) PMID:[17493640](https://pubmed.ncbi.nlm.nih.gov/17493640/)
- Upham BL, Weis LM, Rummel AM *et al.* (1996). The effects of anthracene and methylated anthracenes on gap junctional intercellular communication in rat liver epithelial cells. *Fundam Appl Toxicol*, 34: 260–264. doi:[10.1006/faat.1996.0195](https://doi.org/10.1006/faat.1996.0195) PMID:[8954755](https://pubmed.ncbi.nlm.nih.gov/8954755/)
- Wei S-J, Chang RL, Bhachech N *et al.* (1993). Dose-dependent differences in the profile of mutations induced by (+)-7R,8S-dihydroxy-9S,10R-epoxy-7,8,9,10-tetrahydrobenzo(a)pyrene in the coding region of the hypoxanthine (guanine) phosphoribosyltransferase gene in Chinese hamster V-79 cells. *Cancer Res*, 53: 3294–3301. PMID:[8324741](https://pubmed.ncbi.nlm.nih.gov/8324741/)
- Weis LM, Rummel AM, Masten SJ *et al.* (1998). Bay or baylike regions of polycyclic aromatic hydrocarbons were potent inhibitors of Gap junctional intercellular communication. *Environ Health Perspect*, 106: 17–22. doi:[10.1289/ehp.9810617](https://doi.org/10.1289/ehp.9810617) PMID:[9417772](https://pubmed.ncbi.nlm.nih.gov/9417772/)
- Wenzlaff AS, Cote ML, Bock CH *et al.* (2005a). CYP1A1 and CYP1B1 polymorphisms and risk of lung cancer among never smokers: a population-based study. *Carcinogenesis*, 26: 2207–2212. doi:[10.1093/carcin/bgi191](https://doi.org/10.1093/carcin/bgi191) PMID:[16051642](https://pubmed.ncbi.nlm.nih.gov/16051642/)
- Wenzlaff AS, Cote ML, Bock CH *et al.* (2005b). GSTM1, GSTT1 and GSTP1 polymorphisms, environmental tobacco smoke exposure and risk of lung cancer among never smokers: a population-based study. *Carcinogenesis*, 26: 395–401. doi:[10.1093/carcin/bgh326](https://doi.org/10.1093/carcin/bgh326) PMID:[15528218](https://pubmed.ncbi.nlm.nih.gov/15528218/)
- WHO (1998). Selected Non-heterocyclic Polycyclic Aromatic Hydrocarbons (Environmental Health Criteria 202). Geneva: World Health Organization.
- Whyatt RM, Santella RM, Jedrychowski W *et al.* (1998). Relationship between ambient air pollution and DNA damage in Polish mothers and newborns. *Environ Health Perspect*, 106: Suppl 3: 821–826. PMID:[9646044](https://pubmed.ncbi.nlm.nih.gov/9646044/)
- Yoon HS, Lee KM, Lee KH *et al.* (2012). Polycyclic aromatic hydrocarbon (1-OHPG and 2-naphthol) and oxidative stress (malondialdehyde) biomarkers in urine among Korean adults and children. *Int J Hyg Environ Health*, 215: 458–464. doi:[10.1016/j.ijheh.2012.02.007](https://doi.org/10.1016/j.ijheh.2012.02.007) PMID:[22436105](https://pubmed.ncbi.nlm.nih.gov/22436105/)
- Yu D, Berlin JA, Penning TM, Field J (2002). Reactive oxygen species generated by PAH o-quinones cause change-in-function mutations in p53. *Chem Res Toxicol*, 15: 832–842. doi:[10.1021/tx010177m](https://doi.org/10.1021/tx010177m) PMID:[12067251](https://pubmed.ncbi.nlm.nih.gov/12067251/)
- Zhang Y & Tao S (2009). Global atmospheric emission inventory of polycyclic aromatic hydrocarbons (PAHs) for 2004. *Atmos Environ*, 43: 812–819. doi:[10.1016/j.atmosenv.2008.10.050](https://doi.org/10.1016/j.atmosenv.2008.10.050)
- Zhong Y, Wang J, Carmella SG *et al.* (2011). Metabolism of [D10]phenanthrene to tetraols in smokers for potential lung cancer susceptibility assessment: comparison of oral and inhalation routes of administration. *J Pharmacol Exp Ther*, 338: 353–361. doi:[10.1124/jpet.111.181719](https://doi.org/10.1124/jpet.111.181719) PMID:[21515812](https://pubmed.ncbi.nlm.nih.gov/21515812/)